

Long wavelength data collection

– why & how?



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ESRF Macromolecular
Crystallography Group

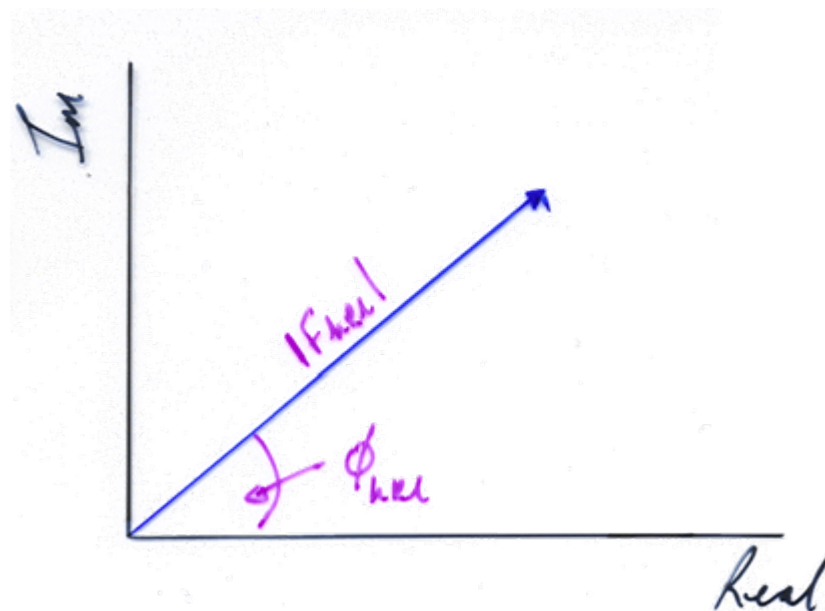
$$\rho_{(x,y,z)} = \frac{1}{V_c} \sum_h \sum_k \sum_l F_{hkl} \exp(-2\pi i(hx + ky + lz))$$

$$F_{hkl} = \sum_j f_j \exp(-2\pi i(hx_j + ky_j + lz_j))$$

$$F_{hkl} = \sum_j |F_{hkl}| \exp(-2\pi i(\phi))$$

↑
Amplitude

↑
Phase



From an x-ray diffraction experiment we can ‘measure’ the amplitude but get *no information about the phase*

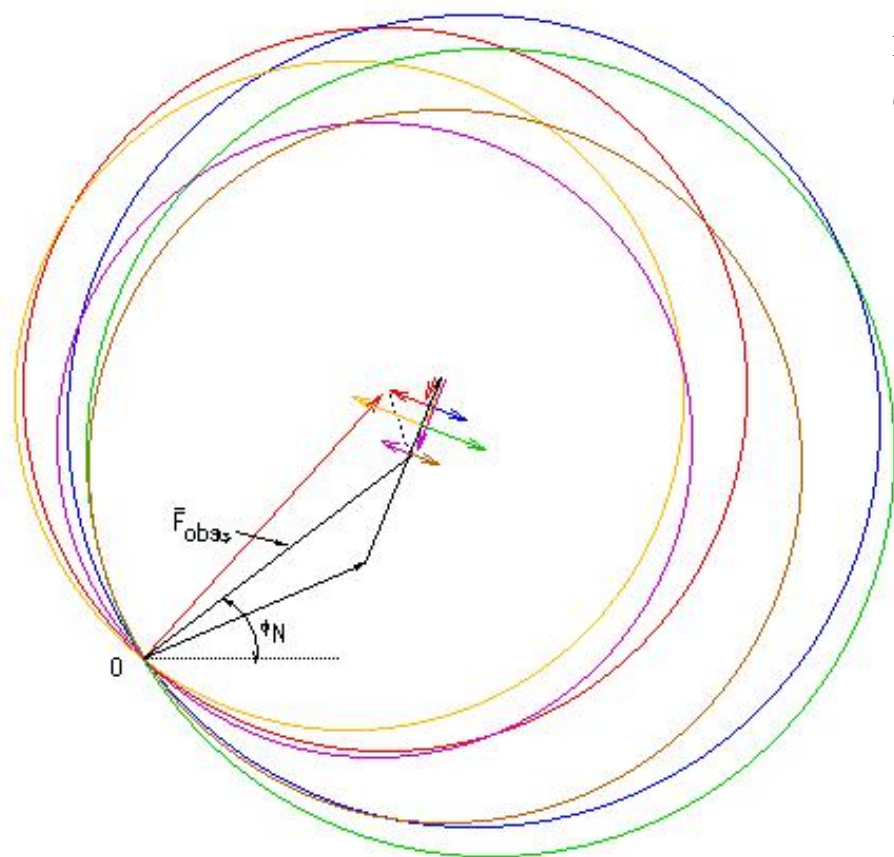
$$F_{hkl} = \sum_j f_j \exp - 2\pi i (hx_j + ky_j + lz_j)$$

$$f = f_o + \underbrace{f'(\lambda) + if''(\lambda)}$$

Wavelength-dependent and vary rapidly
close to an absorption edge

When f'' is large Friedel's Law is no longer obeyed:

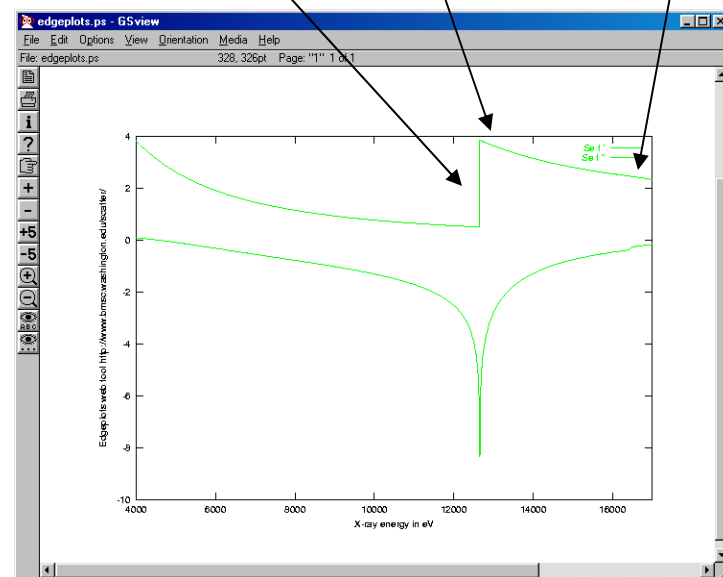
$$F_{hkl} \neq F_{\bar{h}\bar{k}\bar{l}}$$

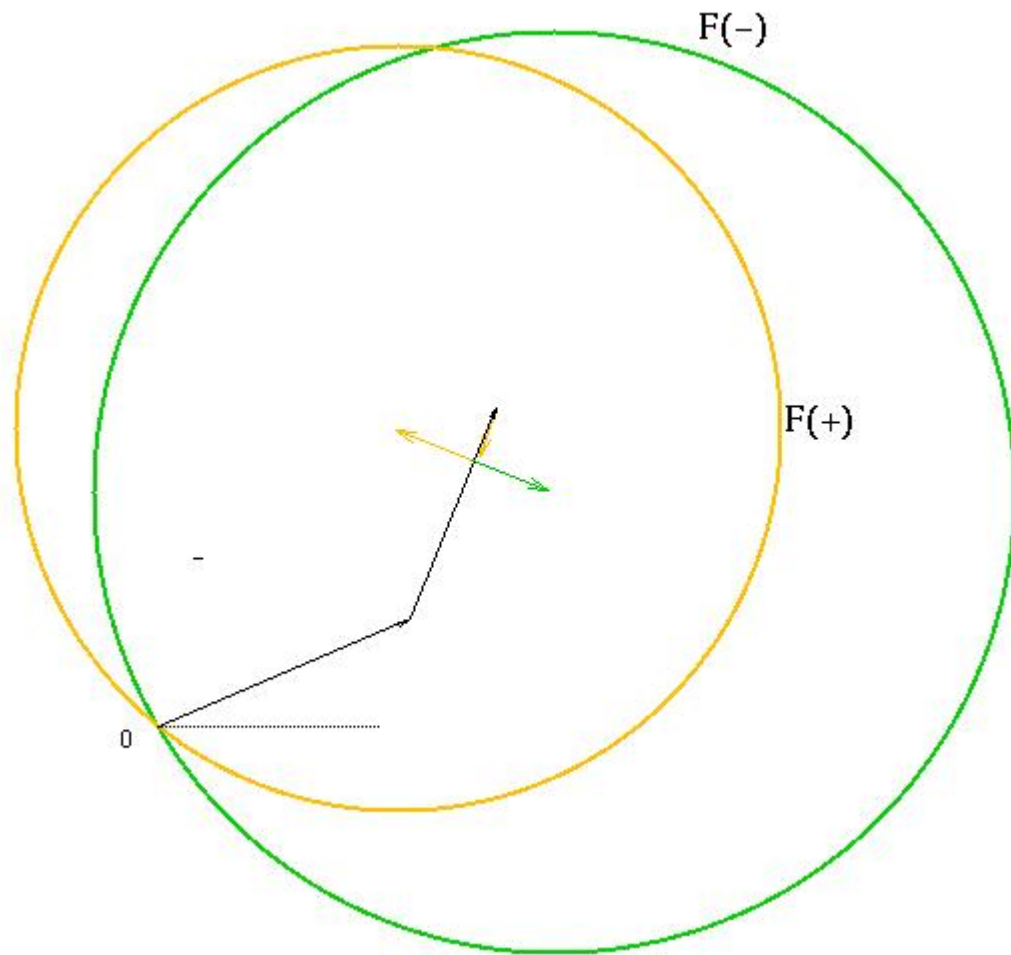


f' at minimum,
moderate value
of f''

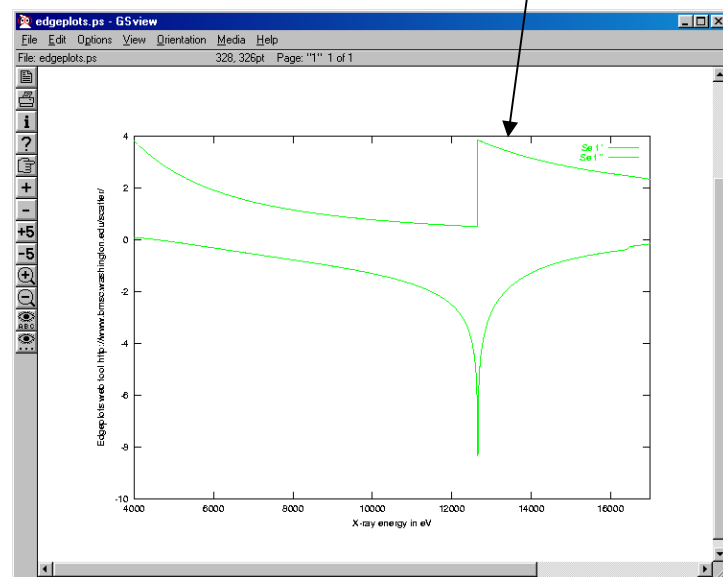
f' moderate,
 f'' large

f' moderate,
 f'' moderate





One can also phase using single wavelength data. Use density modification to break phase ambiguity



K absorption edges: f'' usually has a maximum around $4e^-$
can rise to $6-7e^-$ with ‘white line’.

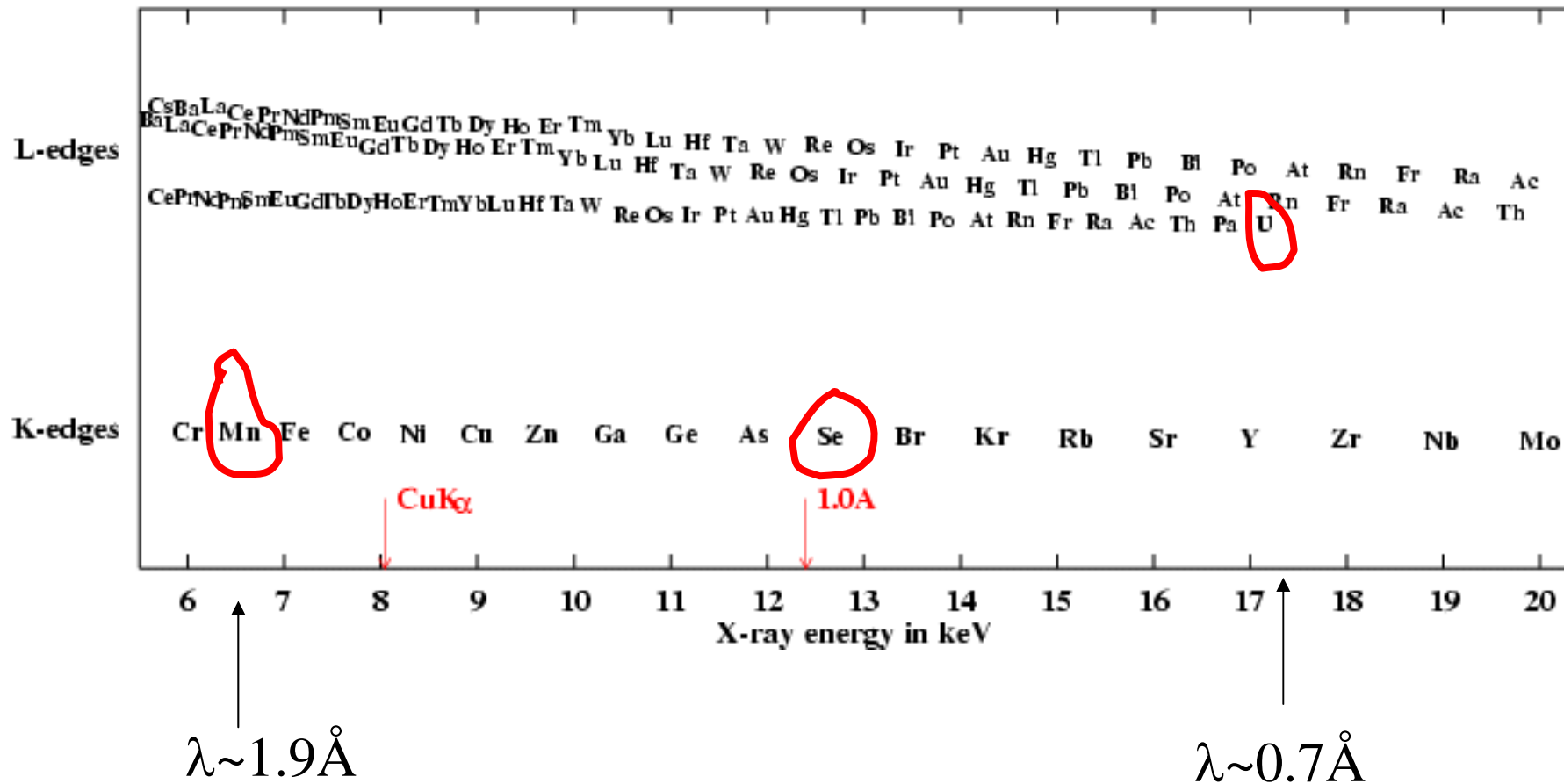
L absorption edges: f'' usually has a maximum around $12e^-$
can rise to $\sim 30e^-$ with ‘white line’.

$$'anomsignal' = \langle \Delta F / F \rangle \approx \frac{1}{\sqrt{2}} \frac{\sqrt{N_A} 2f''}{\langle |F_T| \rangle}$$

“From a practical point of view a larger f'' will give more accurate values for the possible solutions [of the phase]....”

Woolfson & Fan, “Solving Crystal Structures” (1995) Cambridge University Press

Absorption edges useful for anomalous scattering experiments



Are there potential benefits in going beyond this range?

GROUP		Periodic Table of the Elements																VIII											
IA												IIIB		IVB		VB		VIB		VIIB		VIII							
1	2											5	6	7	8	9	10												
3	4											13	14	15	16	17	18												
11	12											21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36		
19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36												
37	38	39	40	41	42	43	44	45	46	47	48	49	50	51	52	53	54												
55	56																	81	82	83	84	85	86						
87	88																	109	110	111	112								
		57	58	59	60	61	62	63	64	65	66	67	68	69	70	71													
		89	90	91	92	93	94	95	96	97	98	99	100	101	102	103													

K-edges of Te, I, Xe accessible at $\sim 0.3 \text{ \AA}$ $\sim (30 \text{ KeV})$

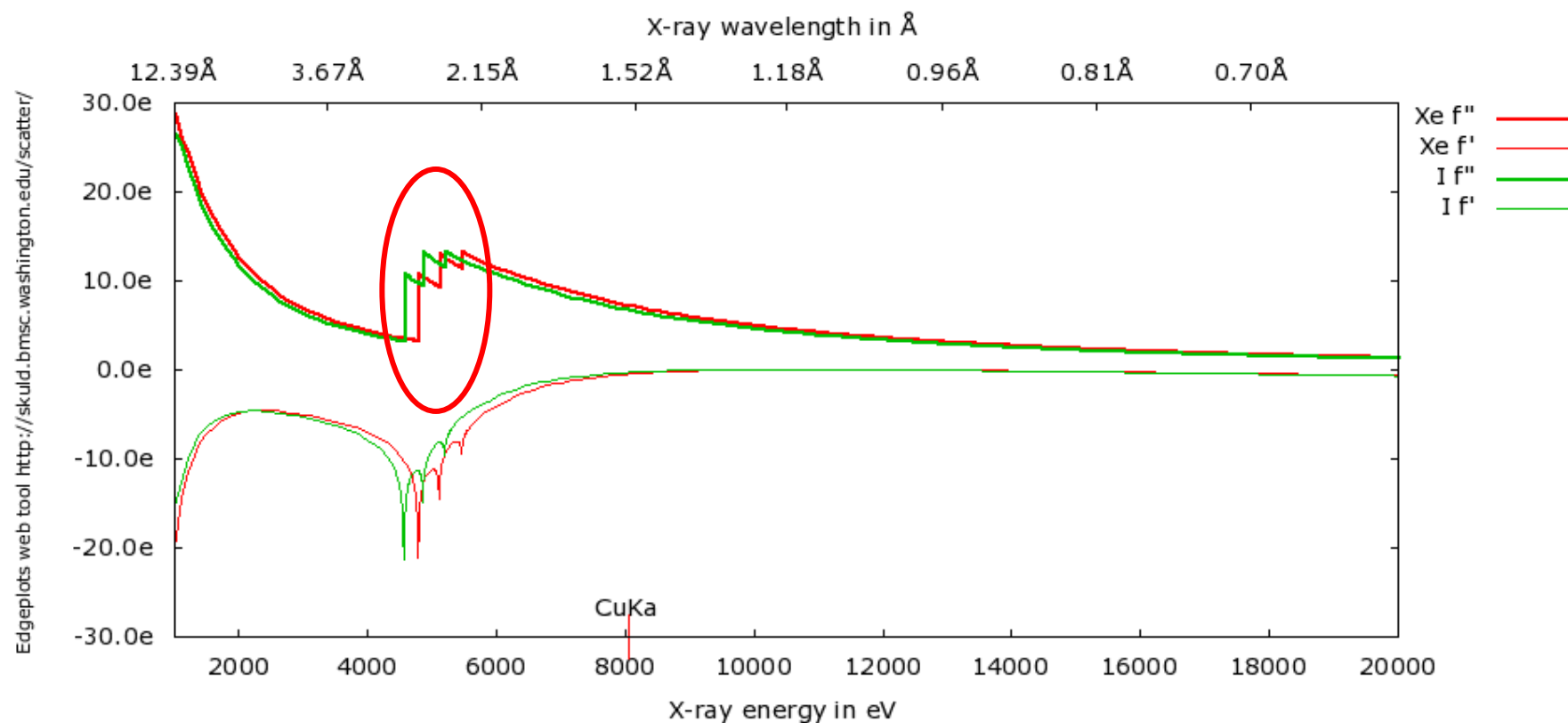
But, K-edges give f'' of (only) $\sim 4 e^-$

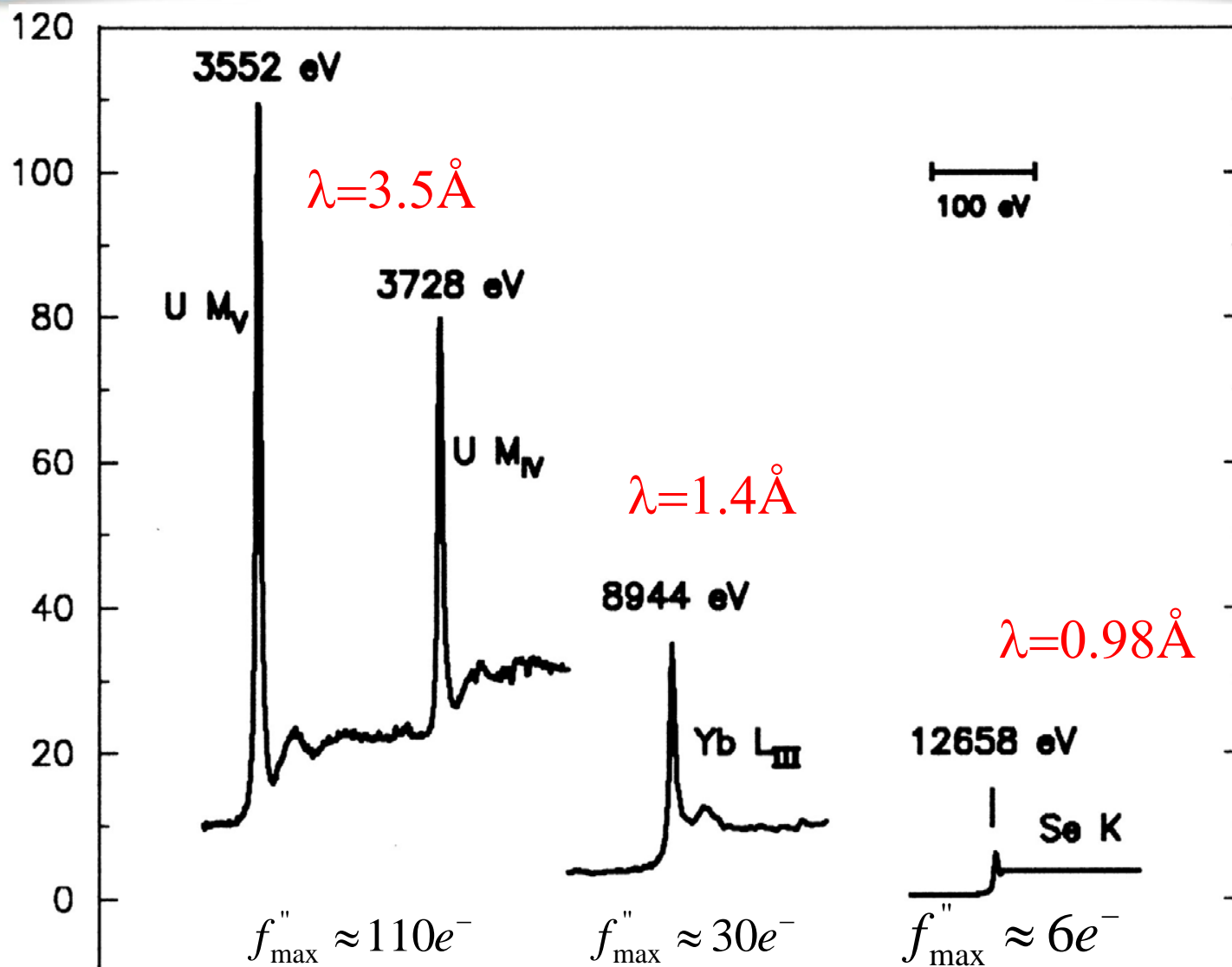
Evans, G. & Bricogne, G. (2002) *Acta Cryst.*, **D58**, 976-991 ; Sauer O., Schmidt A. and Kratky C. (1997) *J. Appl. Crystallogr.*, **30**, 476-486.

Energy (keV)/Wavelength (Å)	$f''_{Xe} \cdot (e)$	$f''_I \cdot (e)$
14.2/0.87	2.8	2.6
12.4/1.00	3.6	3.3
10.0/1.24	5.1	4.8
8.0/1.55	7.4	6.9
7.0/1.78	9.2	8.6
6.5/1.91	10.3	9.7
6.0/2.07	11.6	10.9
5.5/2.25	13.2	12.3

In principle, better to collect data at longer wavelength (even though phasing would be carried out using SAD)

of I & Xe at $\lambda \sim 2.5 \text{ \AA}$



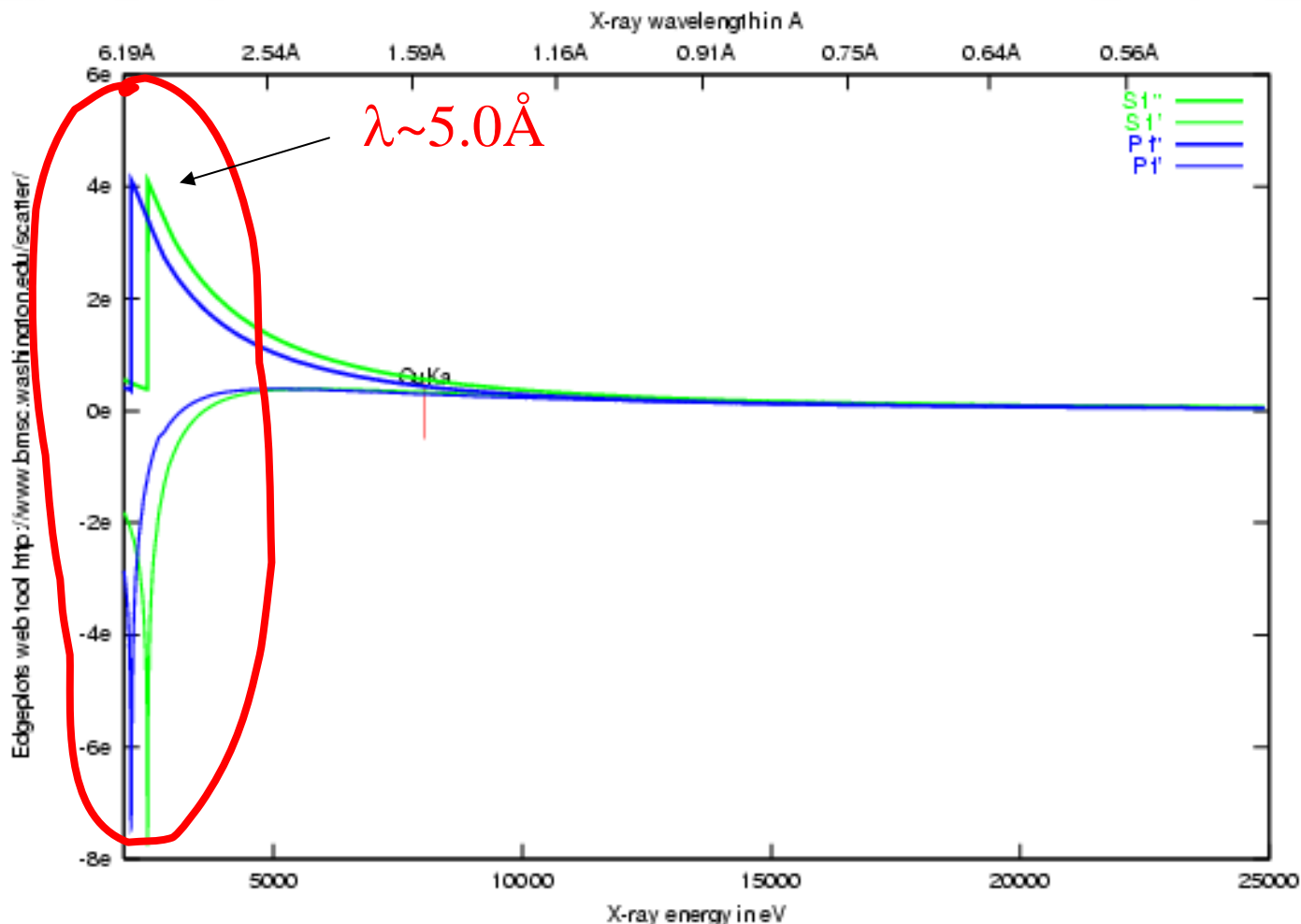


$$\left\langle \frac{\Delta F}{F} \right\rangle \approx \frac{1}{\sqrt{2}} \frac{\sqrt{N_A} 2 f''}{\langle |F_T| \rangle}$$

$$\begin{aligned} \text{For Proteins: } \langle |F_T| \rangle &\sim 6.70 \cdot [\# \text{ Atoms}]^{1/2} \\ &\sim (3.14 \cdot M_r)^{1/2} \end{aligned}$$

**At M_V edge ($\lambda = 3.49 \text{ \AA}$ “ $\sim 110e^-$) $\Delta F/F \sim 6.9\%$ for one U atom in 1.6MDa!!!
(i.e. one fully occupied U atom could ‘phase’ the ribosome!)**

Liu, Ogata, & Hendrickson (2001). Proc. Natl. Acad. Sci. USA, Vol. 98, 10648-10653



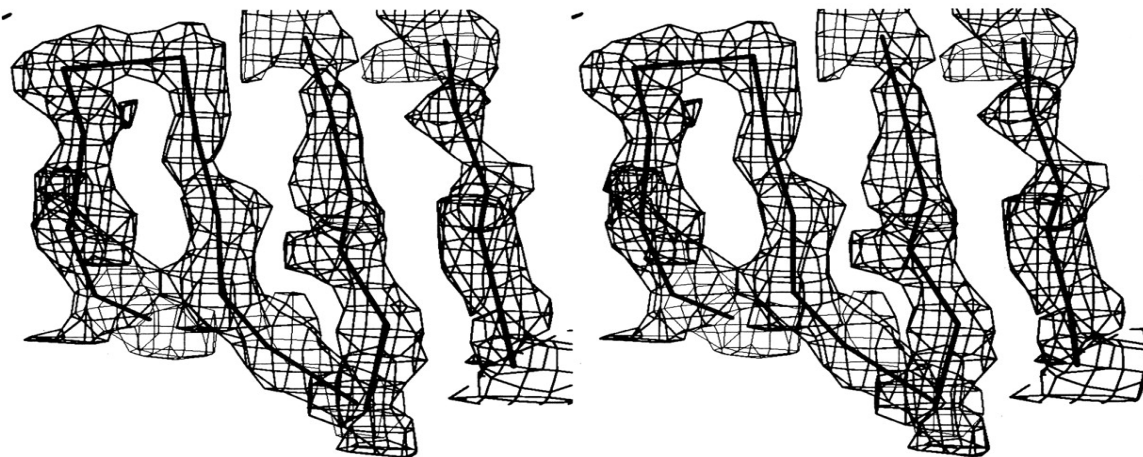
This would provide enough signal to phase the nearly all native macromolecular crystal structures (K-edge of S has a (big) white line)

It is feasible:

Lehmann, Müller & Stuhrmann (1993). *Acta Cryst.* **D49**, 308-310

Kahn, et al. & Stuhrmann (2000). *J. Synchrotron Rad.* **7**, 131-138.

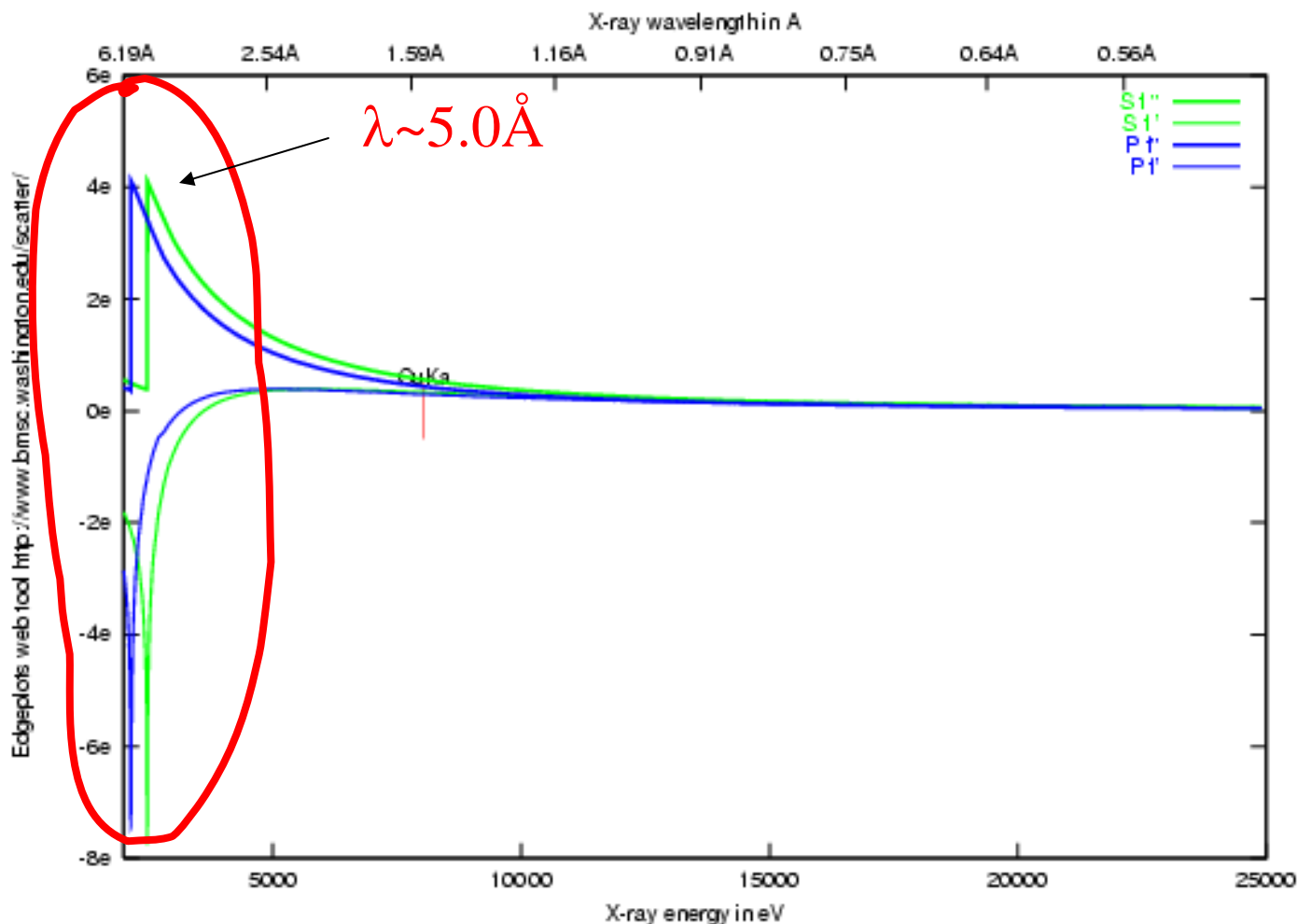
Liu, Ogata, & Hendrickson (2001). *Proc. Natl. Acad. Sci. USA*, **98**, 10648-10653.



Mad-phased electron density
from uranyl derivative of
elastase. Data collected around
U M_{IV} edge

BUT.....

Experiments at S & P K-absorption edges

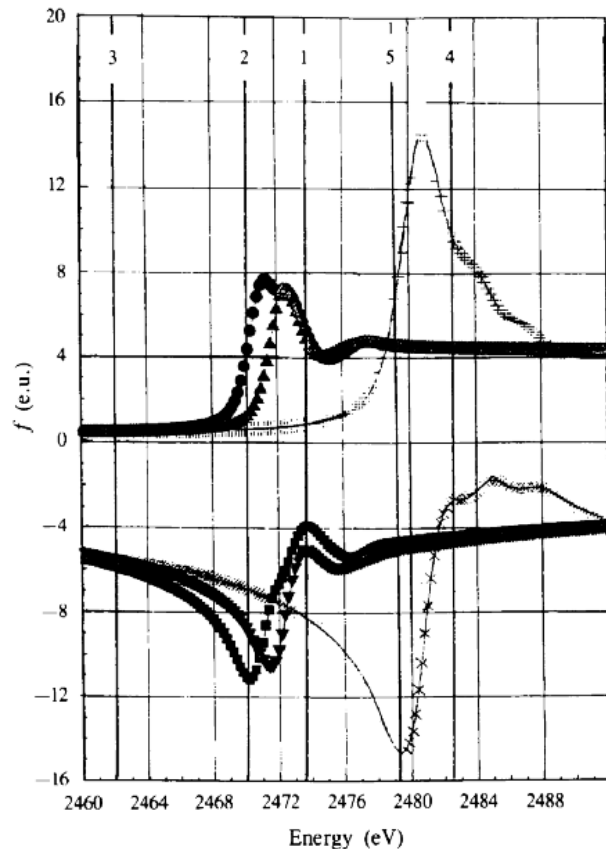


This would provide enough signal to phase the majority of native macromolecular crystal structures



Genome	% S-a.a [#]	$\langle \Delta F/F \rangle$ ($f'' = 4e^-$) (%)
<i>H. sapiens</i>	4.4	6.7
<i>A. thaliana</i>	4.3	6.7
<i>C. elegans</i>	4.7	6.9
<i>D. melanogaster</i>	4.2	6.2
<i>E. coli K12</i>	4.0	6.0
<i>S. cerevisiae</i>	3.4	5.8
<i>C. pneumoniae</i>	3.5	5.8

[#]from <http://www.ebi.ac.uk/proteome/>



White lines will almost double signal!!

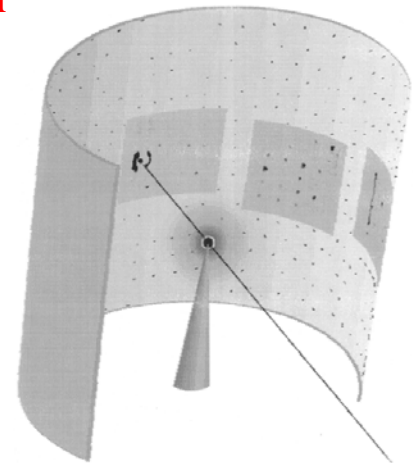
S. Stuhrmann *et al.*, *J. Synchrotron Rad.* (1997). 4, 298-310

Figure 1

Anomalous dispersion of sulfur near the *K*-absorption edge. ▽, △ are f' , f'' of methionine (cysteine); ■, ● are f' , f'' of cystine; × and + are f' , f'' of sulfate ions. The scattering factors are given in electron units (e.u. = scattering length per electron). The numbers inside the figure denote the indices of the chosen energies E_1 – E_5 .

It is experimentally very difficult:

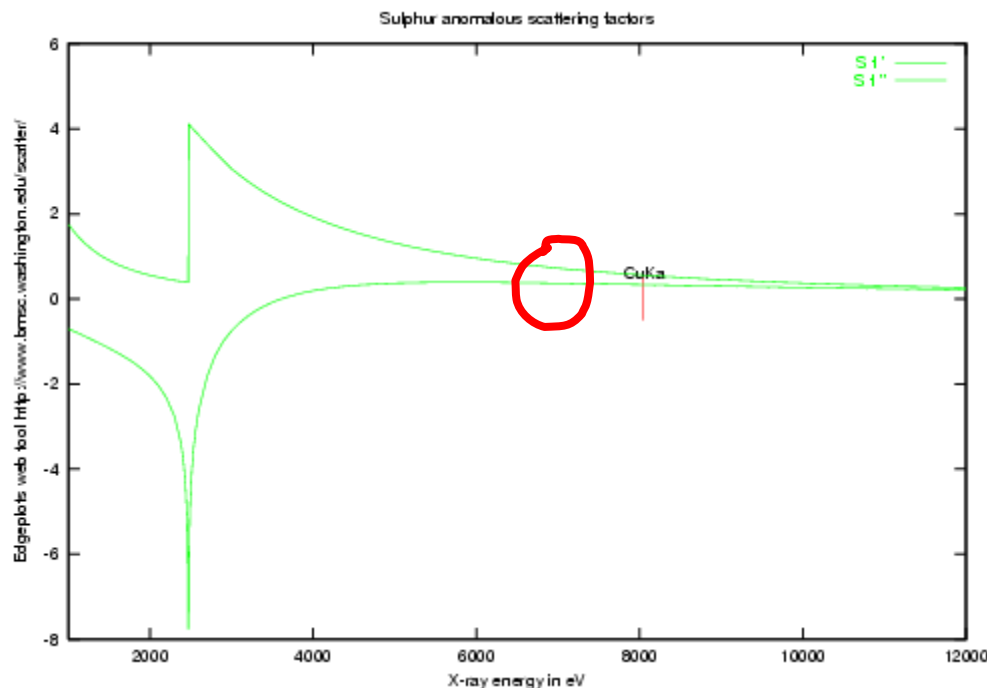
- Absorption (both from air & sample) a real problem. Would need
 - small samples – to reduce absorption
 - evacuated/He-filled ‘experiment’ – reduced air scatter, attenuation of diffracted X-rays
 - specialized beam-lines (no absorbing material between source & sample) – improve intensity at sample position
- Diffraction angles very high
 - $2\theta \sim 112.8^\circ$ at $d_{\min} = 3.0\text{\AA}$ & $\lambda = 5.0\text{\AA}$
 - specially shaped detectors



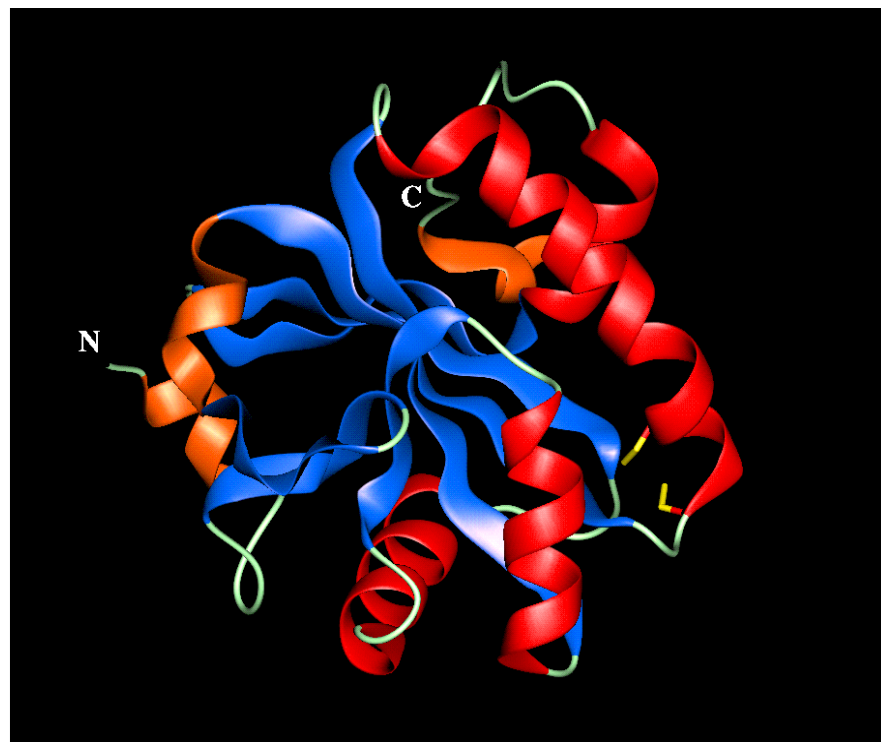
Could probably not do these experiments on a routine basis

- Should we try to exploit this in MX?
- S is naturally present in significant amounts in nearly all proteins.
- No need for heavy atom derivatives (including SeMet)
 - crystal quality not compromised
 - no non-isomorphism
 - less time spent in biochemistry laboratory
 - could truly be a magic bullet

Problems are smaller but so is signal....



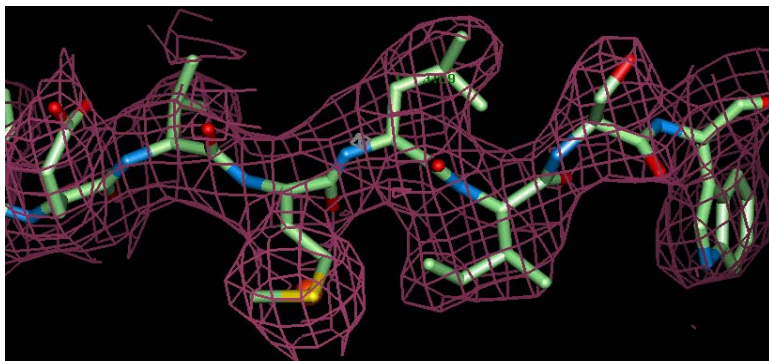
At $\lambda = 1.77\text{\AA}$ $f''=0.72e^-$; Principle already demonstrated by Hendrickson & Teeter (1981), Wang (1985), Dauter (1999).



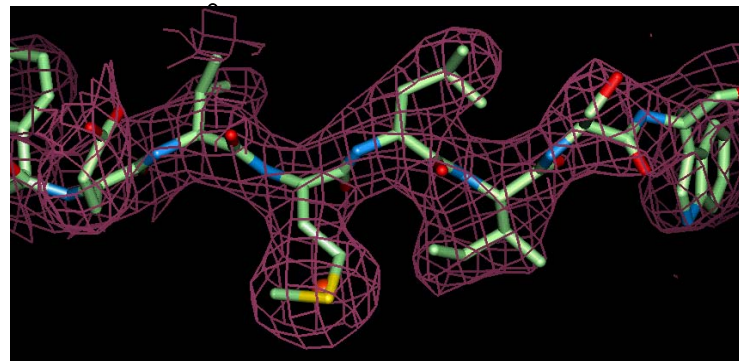
Mol weight	18KDa x 2
<u>Wavelength</u>	<u>1.77Å</u>
Space Group	P4 ₃ 2 ₁ 2
Oscillation range (°)	1.0
No. of frames	560
No. of sulphurs	8 x 2
Resolution range	40 - 2.7Å
Redundancy	30.0
I/σ(I)	52.0
I/σ(I) _{high}	9.5
<u>No. S found</u>	<u>14</u>
<u><ΔF/F></u>	<u>~1.2%*</u>

*Tryparedoxin-II, 14S/300 ordered residues $\langle \Delta F_{hkl} / F_{hkl} \rangle \sim 1.2\%$
 % S-containing residues - 4.7%

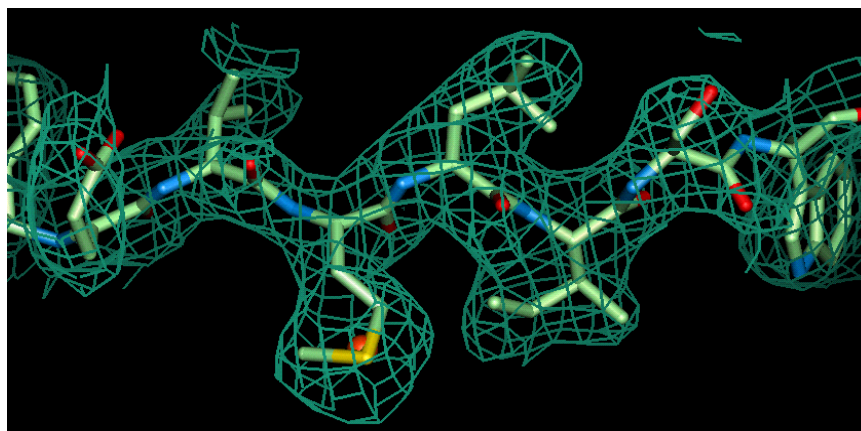
SHARP & Solomon 2.7Å



DM & NCS averaging



DM & NCS averaging 2.35Å



Micossi *et al.*, (2002). *Acta Cryst.* **D58**, 21-28

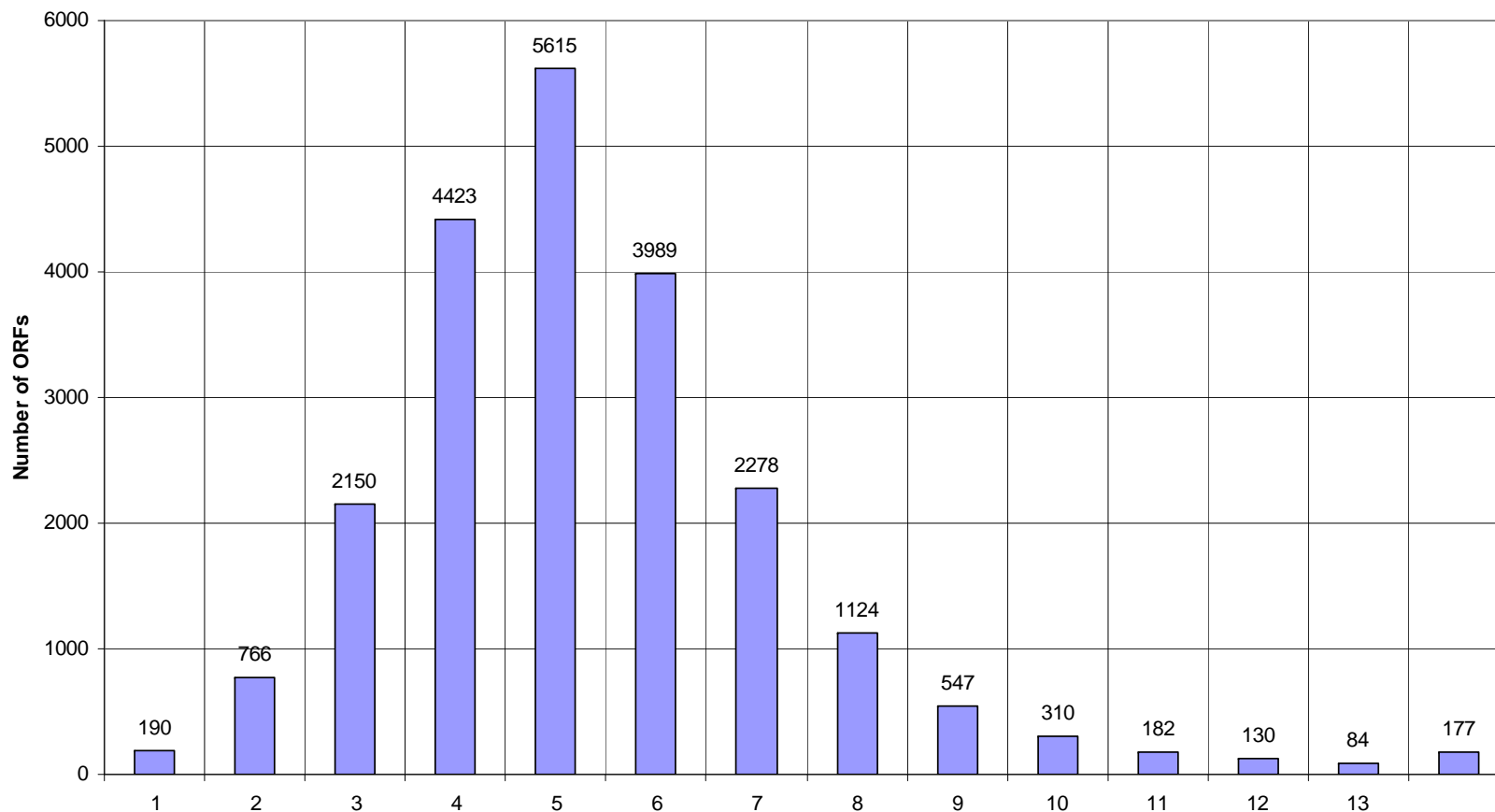


Genome	Met	Cys	Total
<i>Homo sapiens</i>	2.16	2.23	4.39 %
<i>Arabidopsis thaliana</i>	2.45	1.84	4.29 %
<i>Caenorhabditis elegans</i>	2.61	2.06	4.67 %
<i>Drosophila melanogaster</i>	2.35	1.88	4.23 %
<i>Saccharomyces cerevisiae</i>	2.09	1.30	3.39 %
<i>E.coli K12</i>	2.80	1.17	3.97%
<i>Chlamydia pneumoniae</i>	1.92	1.59	3.51%

*Most other Bacteria and Archea have Cys + Met compositions of
between 3-3.5%*

<http://www.ebi.ac.uk/proteome/>

%C+M/ORF in C.elegans



N.B. f'' for S increases at longer wavelengths ($\lambda = 1.9 \text{ \AA}$; $f'' \sim 1.0 \text{ e}^-$)
so we could move to the left of this graph

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Acta Cryst. (2006). D62, 1475-1483 [doi:10.1107/S0907444906038534]

What can be done with a good crystal and an accurate beamline?

J. Wang, M. Dauter and Z. Dauter

Abstract: X-ray single-wavelength anomalous diffraction (SAD) data from a crystal of proteinase K were collected using synchrotron radiation of 0.98 Å wavelength at SER-CAT 22-ID beamline, Advanced Photon Source, Argonne National Laboratory. At this wavelength, the expected Bijvoet ratio resulting from the presence of one calcium, one chloride and ten S atoms in the 279-residue protein is extremely small at ~0.46%. The direct-methods program *SHELXD* located 11 anomalous sites using data truncated to 2 Å resolution. *SHELXE* was used to produce an easily interpretable electron-density map. This study shows that an accurate beamline and a good-quality crystal provide the possibility of successfully using a very weak anomalous signal of sulfur measured at a short wavelength for phasing a protein structure, even if a small degree of radiation damage is present.

PDB reference: 2id8

Keywords: anomalous scattering; SAD phasing.

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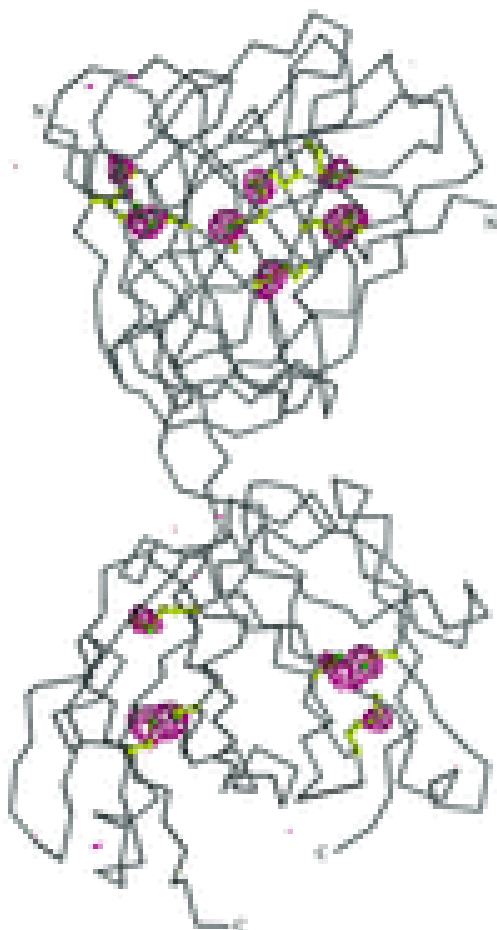
Acta Cryst. (2003). D59, 1020-1027 [doi:10.1107/S0907444903007467]

Phasing on anomalous signal of sulfurs: what is the limit?

U. A. Ramagopal, M. Dauter and Z. Dauter

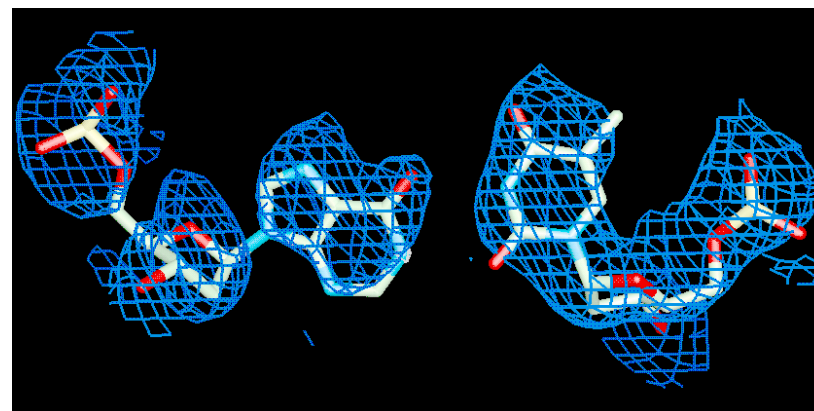
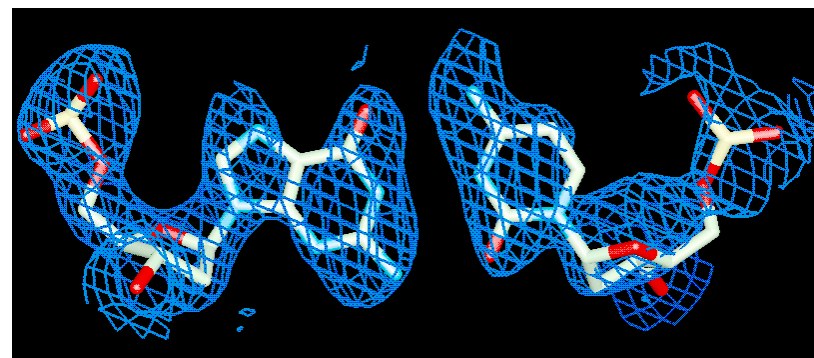
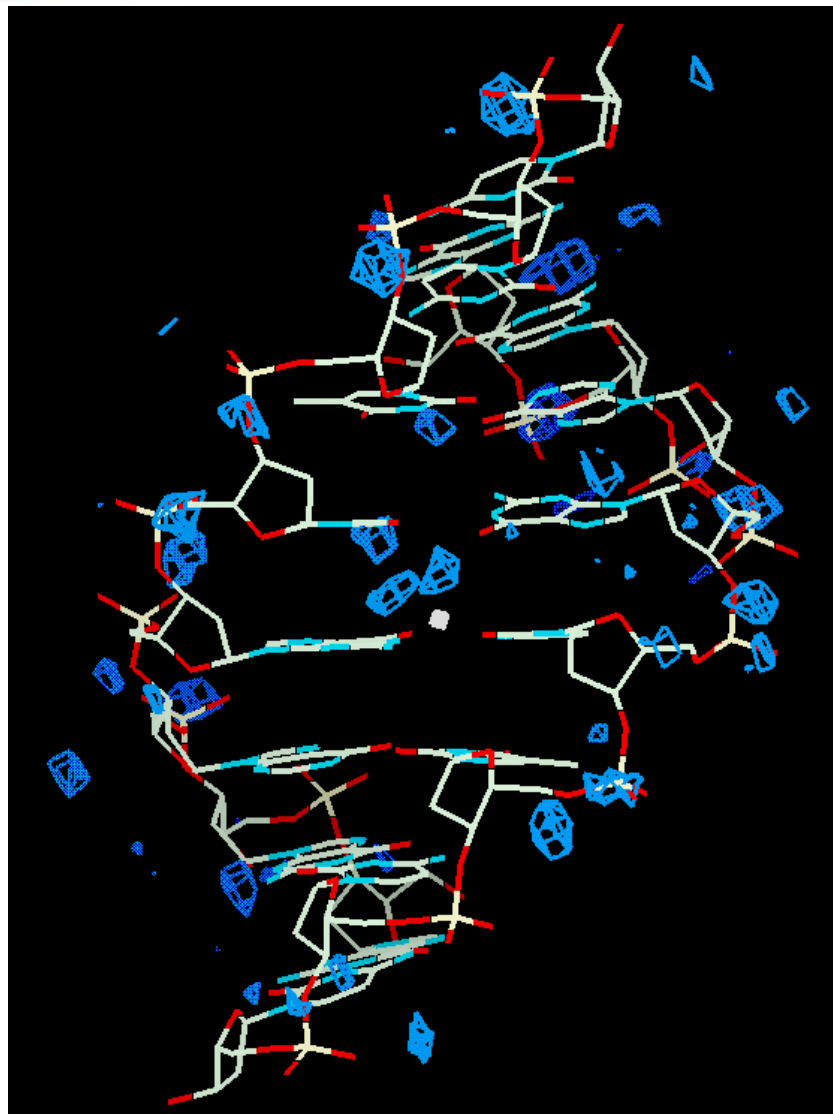
Abstract: Recent years have witnessed significant advancements in X-ray data-acquisition techniques and phasing algorithms, which have made possible the successful use of a very small anomalous diffraction signal for the solution of crystal structures of macromolecules. Two crystal structures, a 44 kDa glucose isomerase containing nine sulfurs and a 33 kDa xylanase containing five sulfurs, have been solved from single-wavelength anomalous data using widely available methods and programs. These two enzymes contain less sulfur than most proteins in the bacterial or eukaryotic proteomes, providing a Bijvoet ratio of about 0.5%. For glucose isomerase the automatically interpretable electron-density maps could be obtained at high as well as low resolution. The S-SAD approach relies on the anomalous signal of sulfur naturally occurring in proteins and alleviates all need for sample derivatization. It may therefore be applicable to all protein crystals able to provide accurate diffraction data.

Keywords: anomalous scattering; sulfur; SAD; xylanase; glucose isomerase.

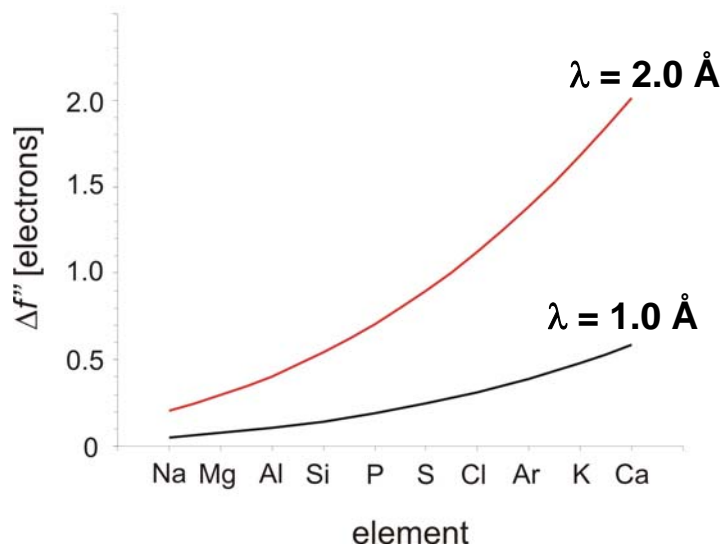


Anomalous difference fourier map (ΔF_{ano} , $\alpha - 90^\circ$) calculated using phases from a MR solution and anomalous differences measured at $\lambda = 1.54 \text{ \AA}$. The peaks in the map represent the 15 S atoms in the asymmetric unit (Schuermann & Tanner, (2003) *Acta Crystallogr. D59*, 1731-1736).

Two choices: 1) Use S positions as makers in map interpretation. 2) Calculate phase probability distributions using anomalous differences & S positions. Use these to get an *unbiased* electron density map.



Maps resulting from SHARP phases subject to density modification with DM. $d_{\min} = 2.4\text{\AA}$
 Model: unrefined MR solution using data to $d_{\min}=3.0\text{\AA}$



Anomalous difference fourier maps ($\Delta F_{\text{ano}}, \alpha - 90^\circ$) will reveal which 'waters' are really ions. Will stand-out better if data collected at longer wavelength. Use XRF analysis to find which ions are likely to be present. If more than one type of ion, peak heights (and coordination sphere) will help discriminate.

Mueller-Dieckmann, C., *et al.*, (2007). *Acta Cryst.*, **D63**, 366-380.

Use what you've learned today – it all applies (probably even more) at long λ !!

Other measures you can take include:

- Reduce air scatter/absorption of diffracted beams
 - He-filled sample environment? vacuum until end of slit box?
- Eliminate $\lambda/3$ contamination of beam by harmonics from monochromator
 - better use of pushers, use of small mirror before sample
- Routine use of (mini-)kappa (**see talk by A. McCarthy**)
 - Anomalous differences measured on same image, same X-ray dose
 - Collection about more than one axis
- *Correcting for/reducing absorption by sample (crystal + surroundings)
 - Analytical absorption corrections?
 - 'loopless' mounting of frozen crystals
 - Scaling long wavelength data against reference data set?
- Better treatment & use of radiation damage (**see talk by M. Weik**)
- Use software to help predict strategy & reduce radiation damage (**see talk by A. Popov**)

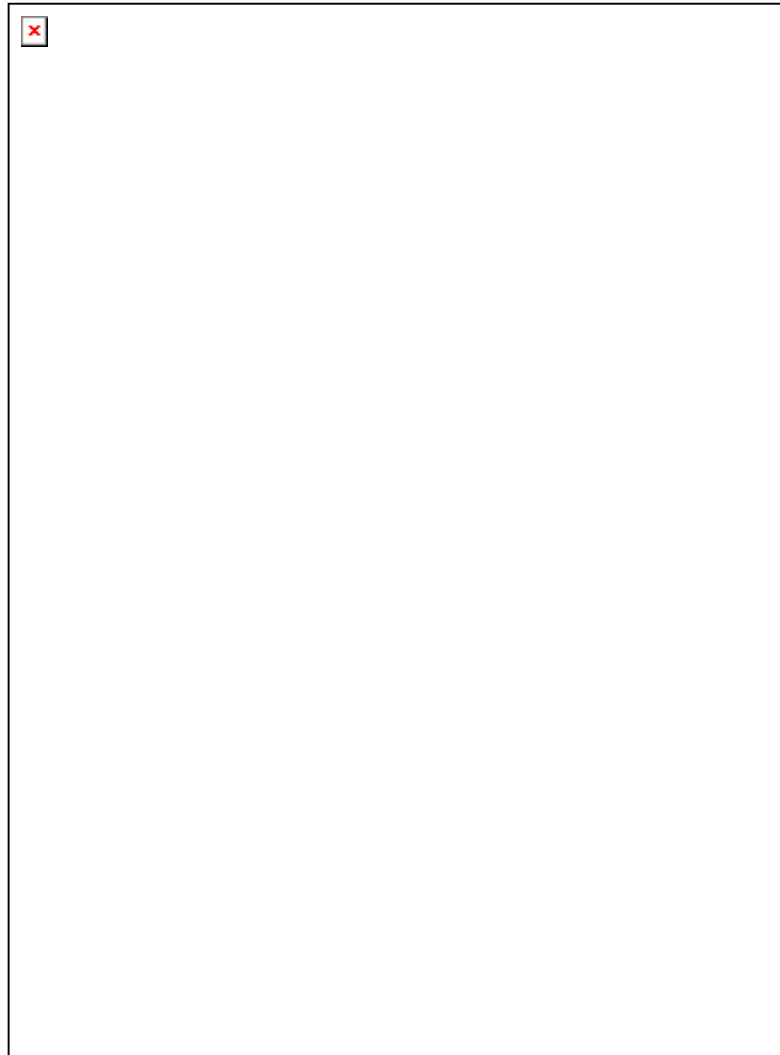
Increase the signal

- **Is there an optimum wavelength for collecting our data? Is it $\lambda = 2.1 \text{ \AA}$? (S-SAD)

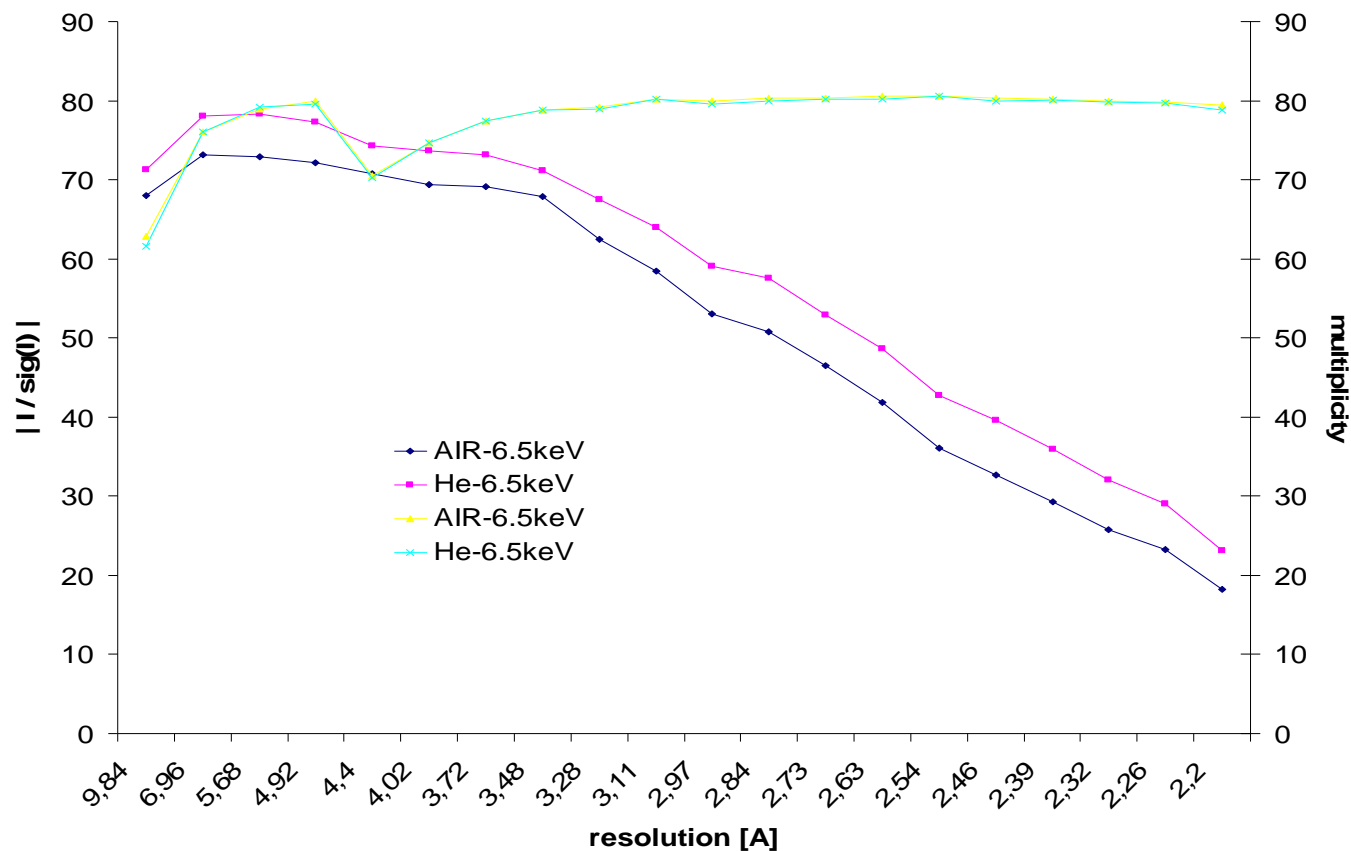
Make sure we collect enough data

- Automatic substructure determination (and more, **see talks by T. Schneider & S. Panjikar**)

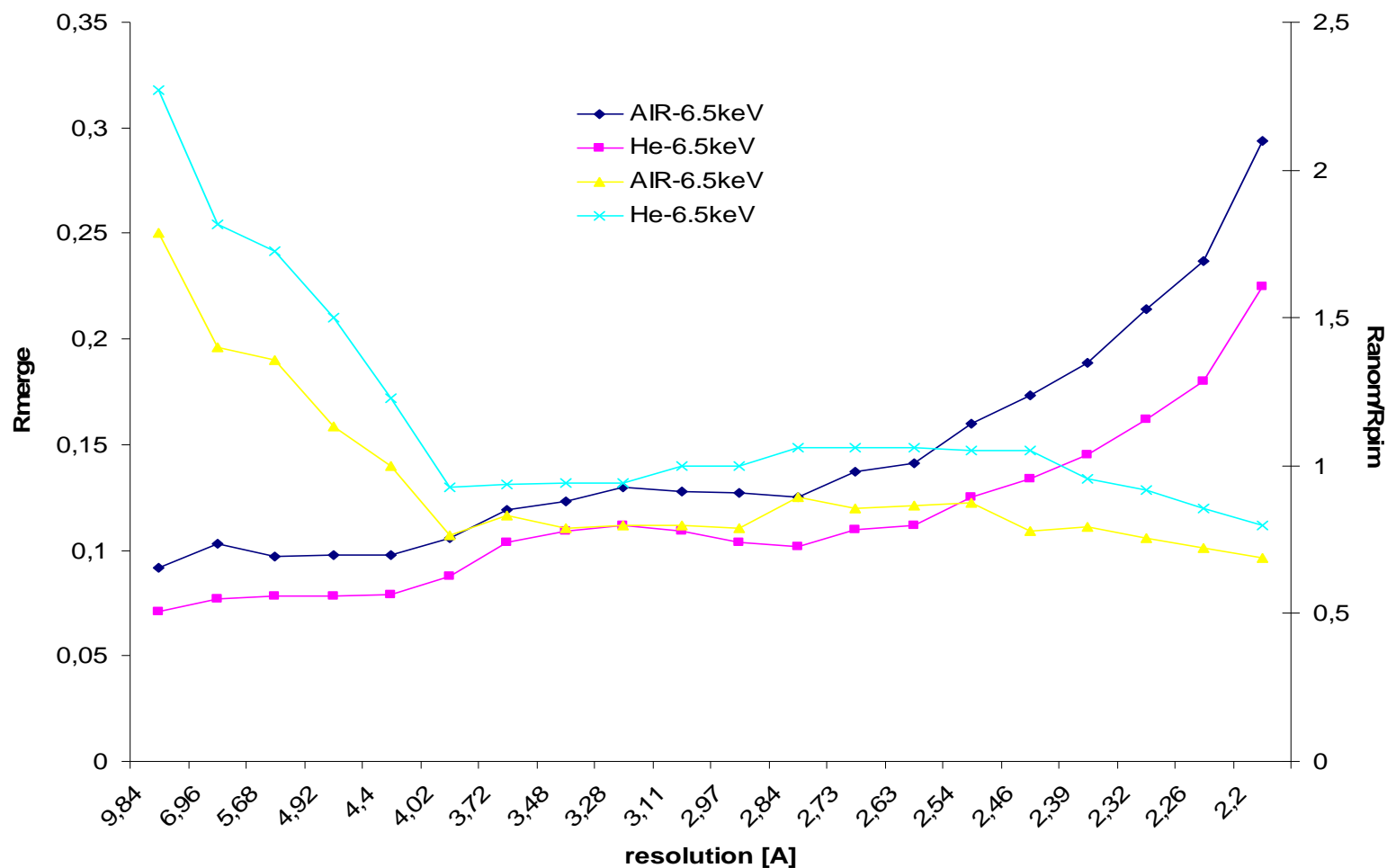
*Brockhauser *et al.*, (2008). *J. Appl. Cryst.*, **41**, 1057-1066; Leal, *et al.*, (2008) *J. Appl. Cryst.*, **41**, 729-737. **Mueller-Dieckmann *et al.*, (2004) *Acta Crystallogr.* **D60**, 28-38.



Thermolysin



Termolysin



Anomalous Scattering Differences - Microsoft Internet Explorer

File Edit View Favorites Tools Help

Address http://www.ruppweb.org/new_comp/anomalous_scattering.htm

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Anomalous Scattering Ratios

Anomalous scatterer (element symbol) :

How many anomalous scatterers in molecule :

Number of residues in molecule :

Edge used for MAD experiment : with following energies used for calculation :

(1) XrayAnode ,

(2) Edge minus , (3) Edge minus , (4) Edge plus , (5) Edge plus eV

Wavelength/Energy Conversion

Enter either a wavelength (shorter than 100 Å) or an energy (larger than 100 eV) :

or select an anode material

Internet

Expected anomalous dispersion ratios - Microsoft Internet Explorer

Address: <http://www.ruppweb.org/cgi-bin/noweb.exe?Element=5&iano=4&nresi=100&iwavl=K&w1=NONE&w2=1000&w3=2&w4=1&w5=4500&anoweb=Submit>

Running on www.ruppweb.org/cgi-bin/xsect.dat

Anomalous scatterer s, z = 16
 K-edge 2472.00eV
 L1-edge 229.20eV
 L2-edge 164.80eV
 L3-edge 164.80eV

4 anomalous scatterers (s) per molecule
 100 residues in molecule, estimated number of protein atoms is 810.
 Estimated Z(eff) of 6.70, q-factor is 0.00742. Calculating.....

Energy (eV), Wavelength (Å) and scattering factors used for S K -edge:

Energy	Wavelength	f'	f''
1472.00	8.4223089	-1.1316	0.9287
2470.00	5.0192871	-7.7604	0.3836
2473.00	5.01320	-8.3768	0.1071
6972.00	1.7782041	0.3693	0.7249

The diagonal elements of the following matrix contain the Bijvoet ratios (i.e. for Friedel opposites at the same wavelength), and the off-diagonal elements the dispersive ratios between different wavelengths. To obtain a usable dispersive signal, the data must be measured with a significantly better (lower) noise level, which can be determined by deriving the Bijvoet ratios from measured centric reflections. [Click here for more explanation.](#)

	1472.00	2470.00	2473.00	6972.00
1472.00	8.42231	5.01929	5.01320	1.77820
2470.00	0.014	0.049	0.055	0.011
2473.00	0.006	0.006	0.066	0.060
6972.00	0.061	0.011	0.011	0.011

Using a longer wavelength will increase signal.....

Thanks for your attention!