

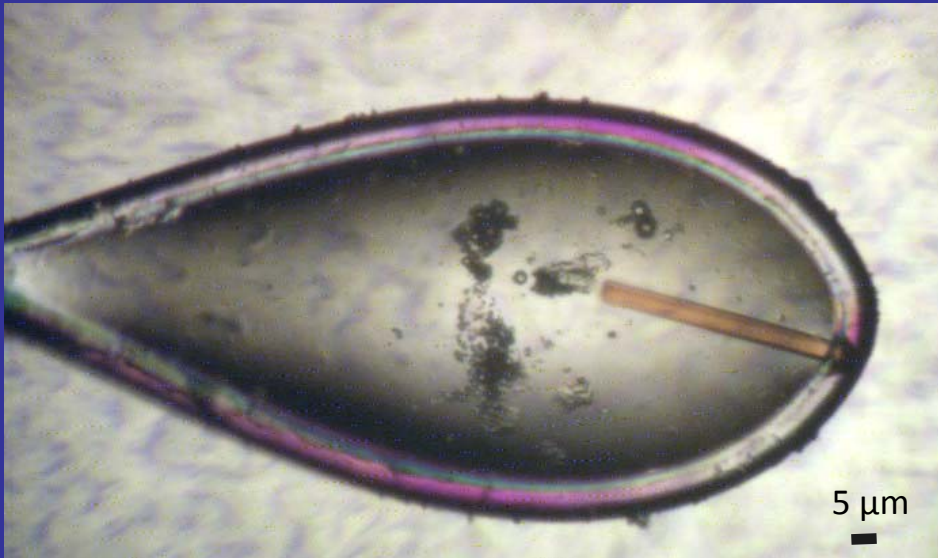
Collecting data on Microfocus beam lines

Pat Edwards

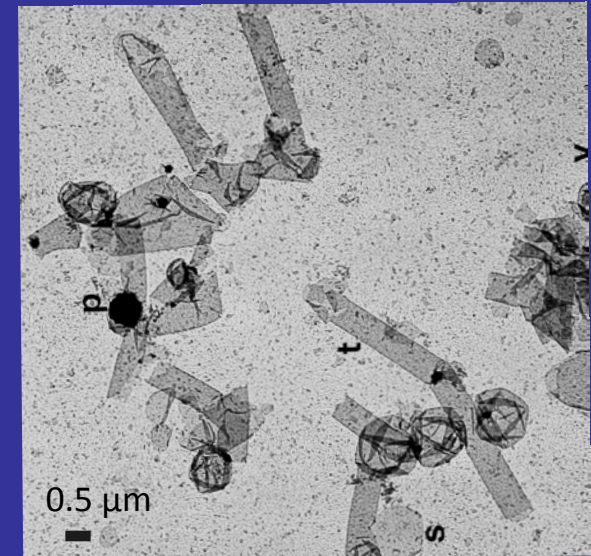
Schertler Group
Structural Studies
MRC Laboratory of Molecular Biology
Cambridge

Why microfocus?

Rhodopsin; 3D and 2D crystal comparison



3D



2D

Microfocus is useful for difficult targets

Membrane proteins; our targets are eukaryotic

- difficult to express
- low yield
- difficult to purify, need detergents and lipids
- difficult to crystallise
- lipid cubic phase

But crystallisation robotics now give more chances
with less protein

Microfocus is useful for difficult targets

Rhodopsin;

- smaller crystals diffracted better than large ones
- first good diffraction found on ID13 in 1997
- general tendency for thin needles, anisotropic in needle direction



Collecting data on Microfocus beam lines

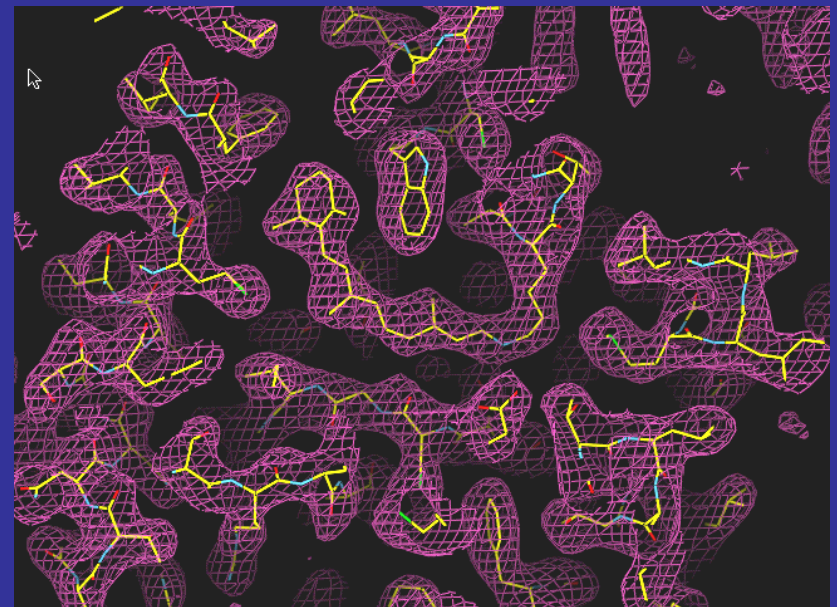
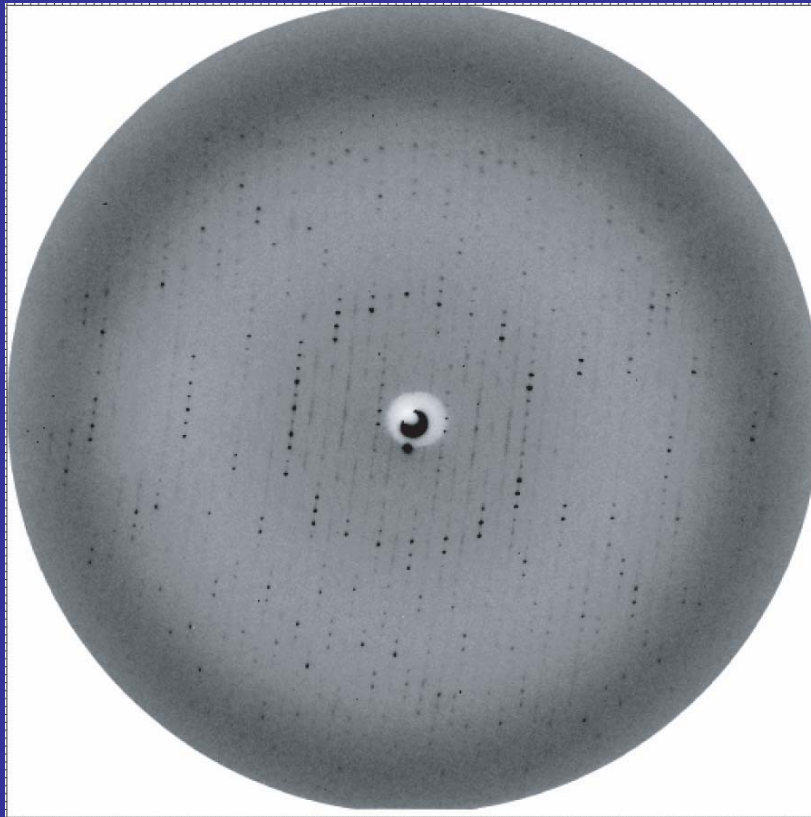
ID13; Manfred Burghammer, Christian Riekel

ID23-2; David Flot, Sean McSweeney



The GPCR bovine rhodopsin - 1999

first good diffraction with 10 μ m beam



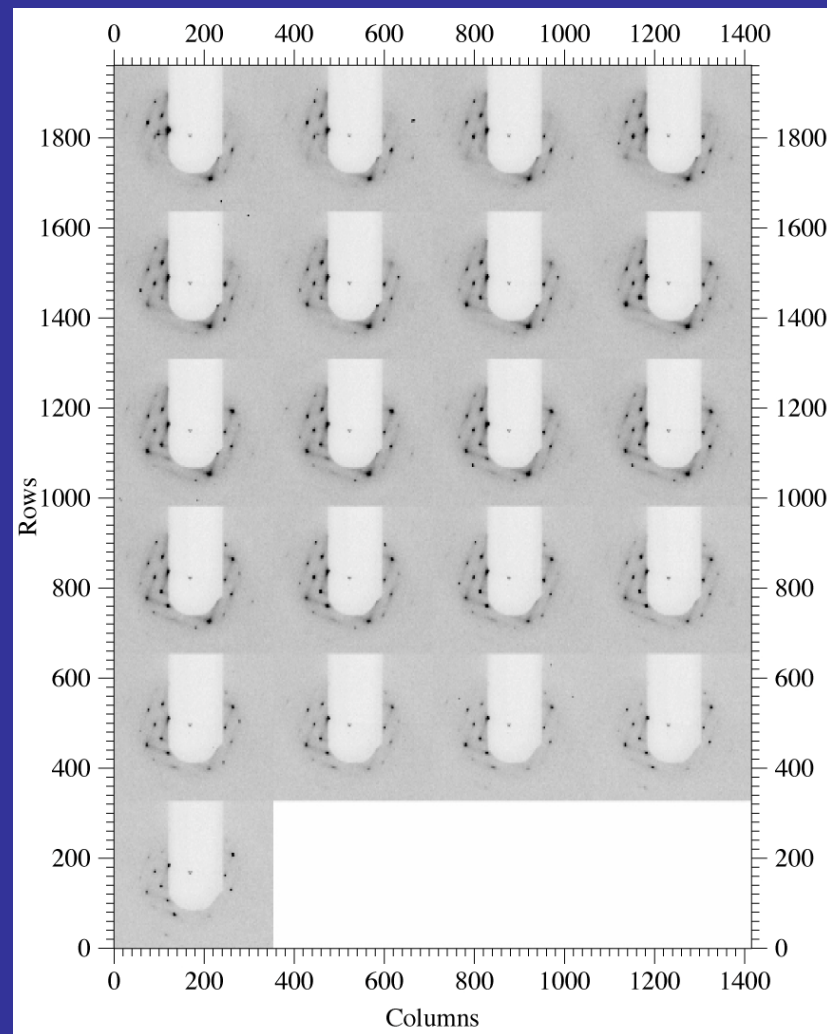
Li *et al*, JMB 2004



Human beta2 adrenoceptor micro crystal

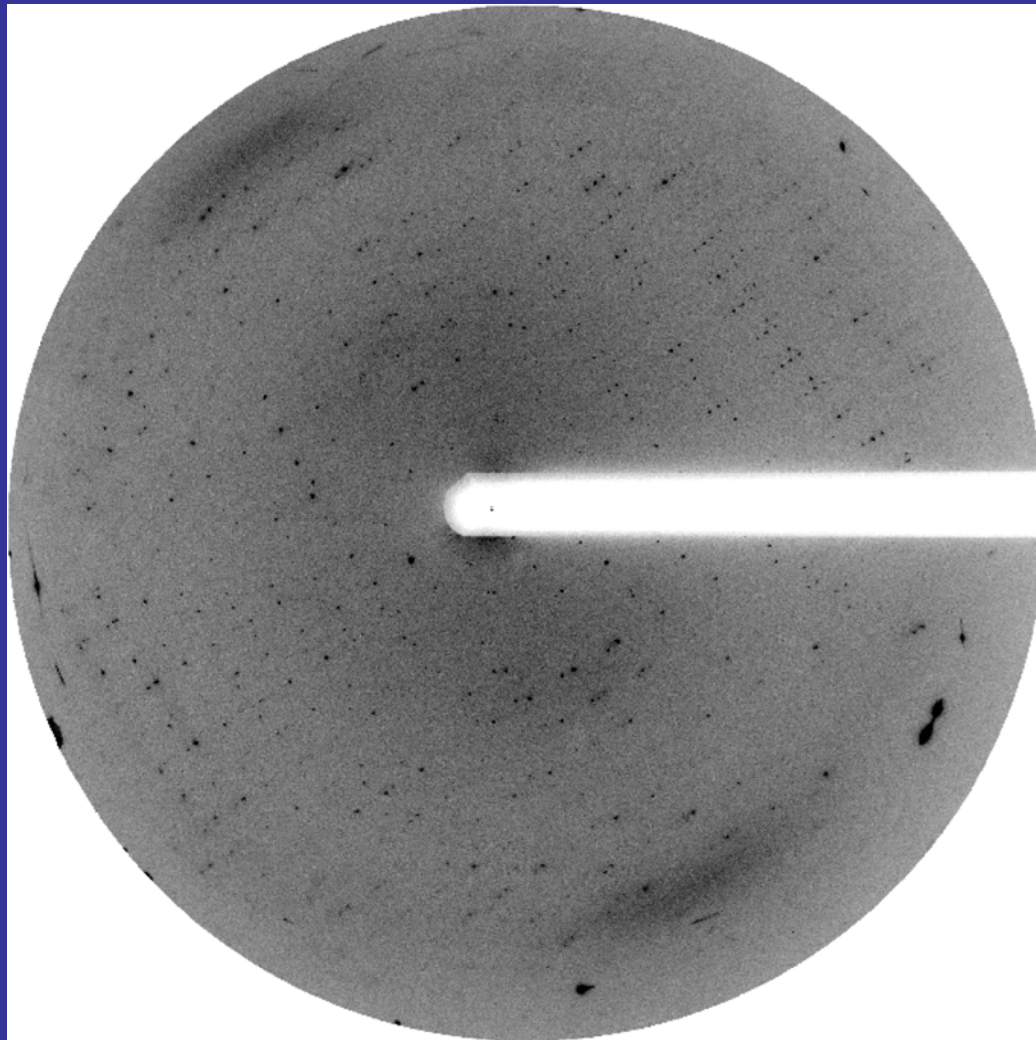
ID13 5 μ m beam

Initial hits

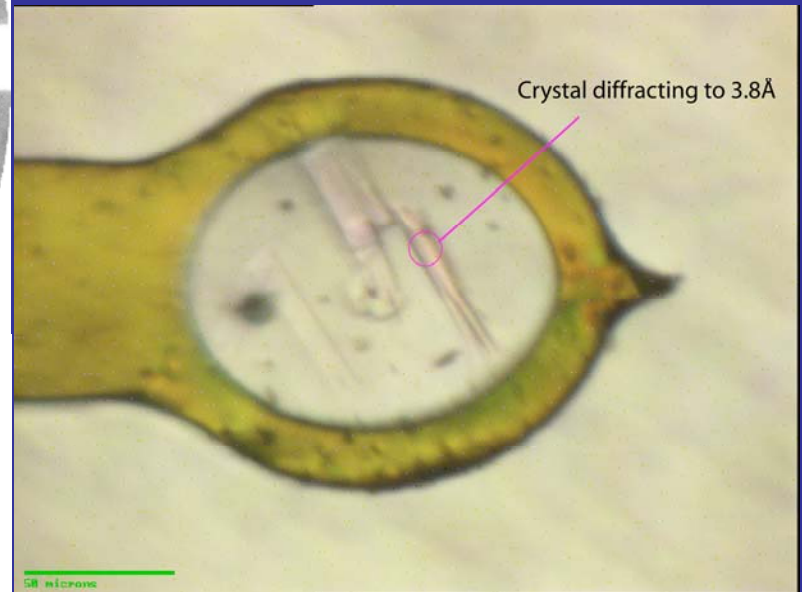




FAB-beta2 complex



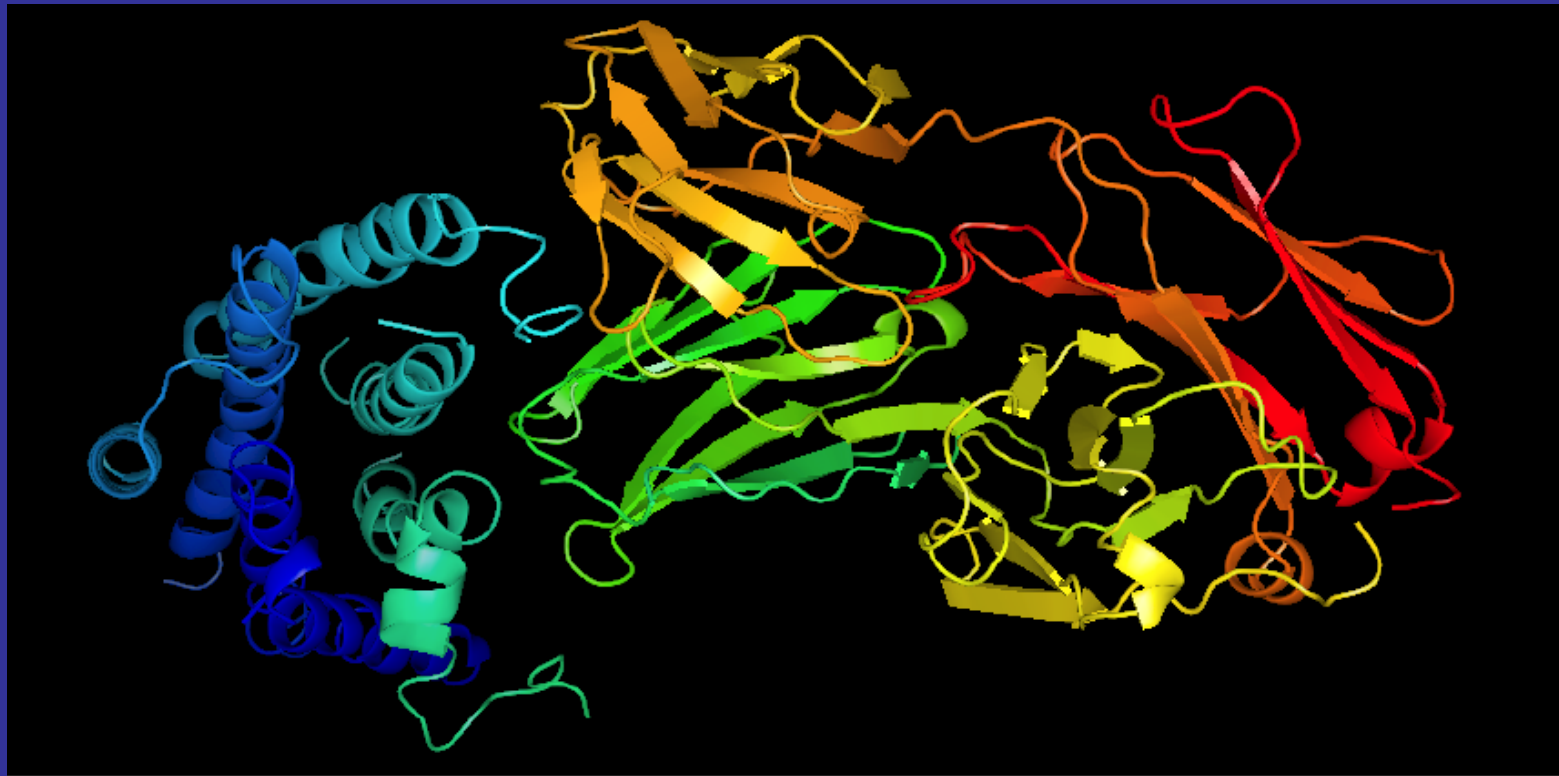
Diffraction is slightly better than 3.8 Å



Fab-beta 2 crystal in loop



Structure of human beta2 in complex with the antibody fragment

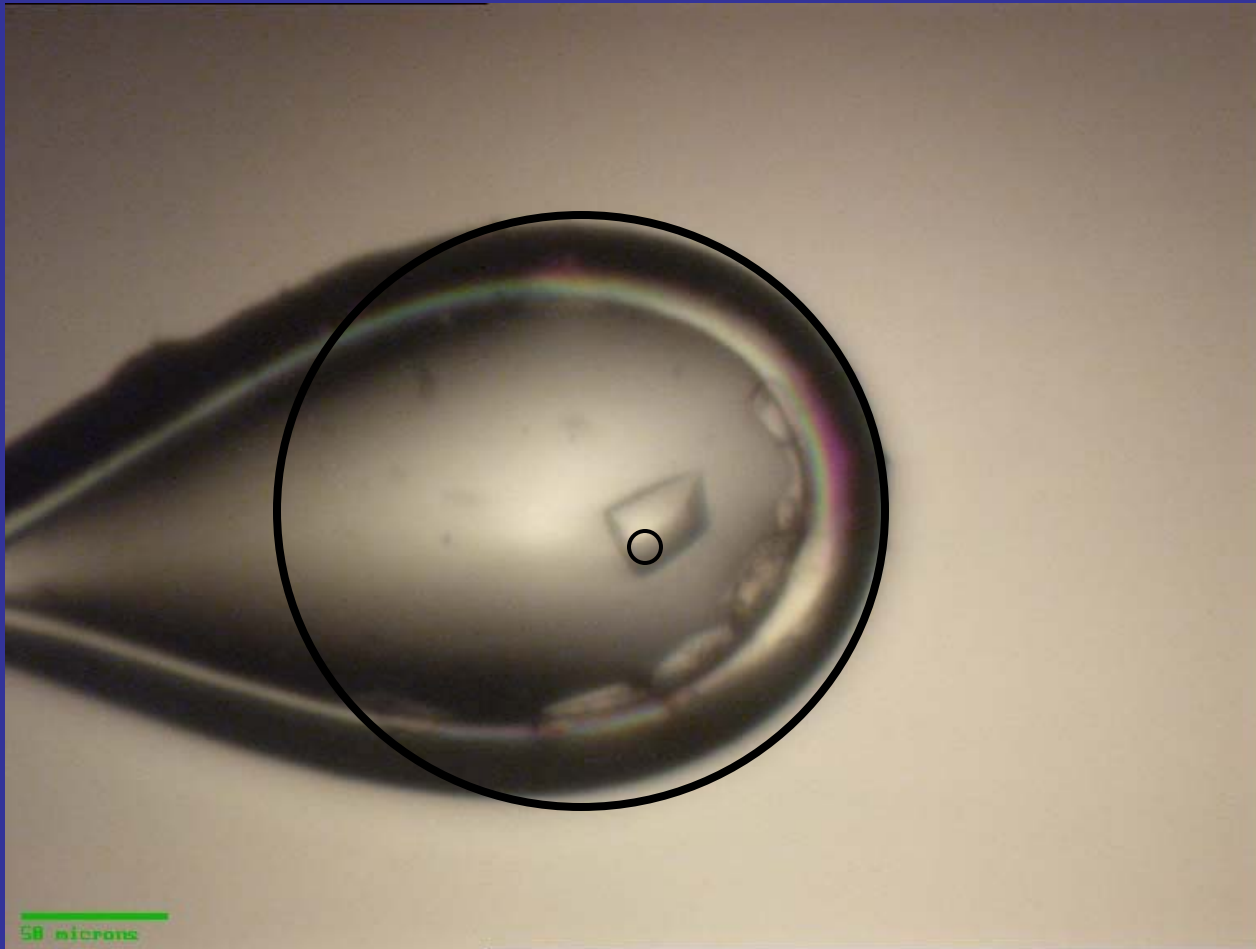


Rasmussen *et al*, Nature 2007



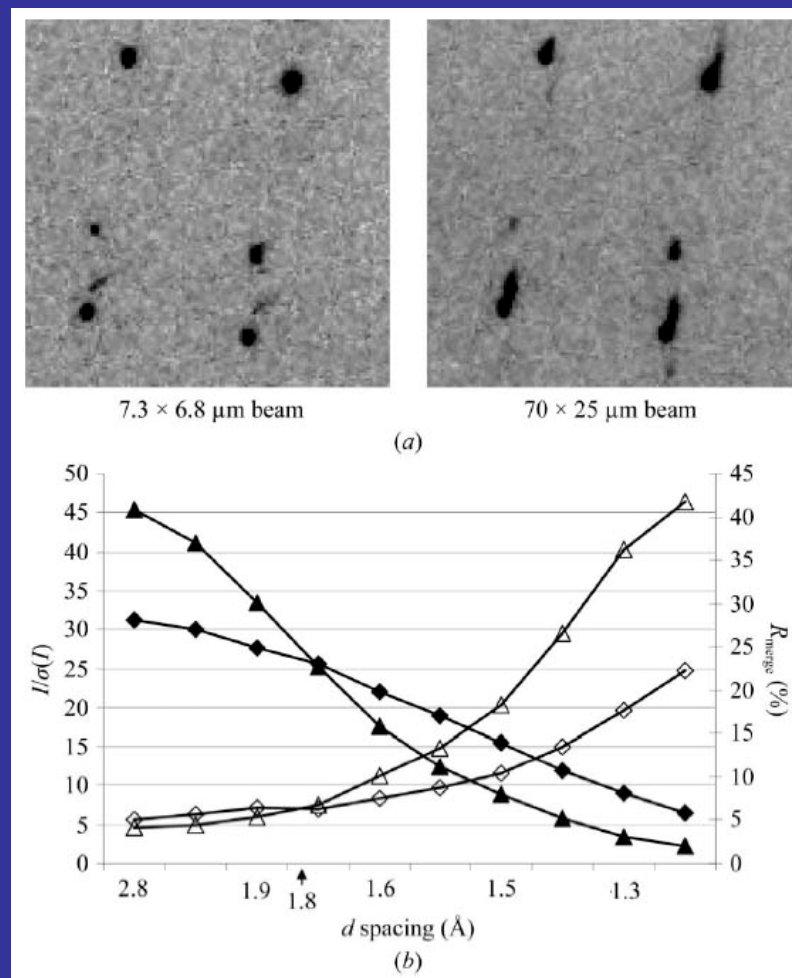
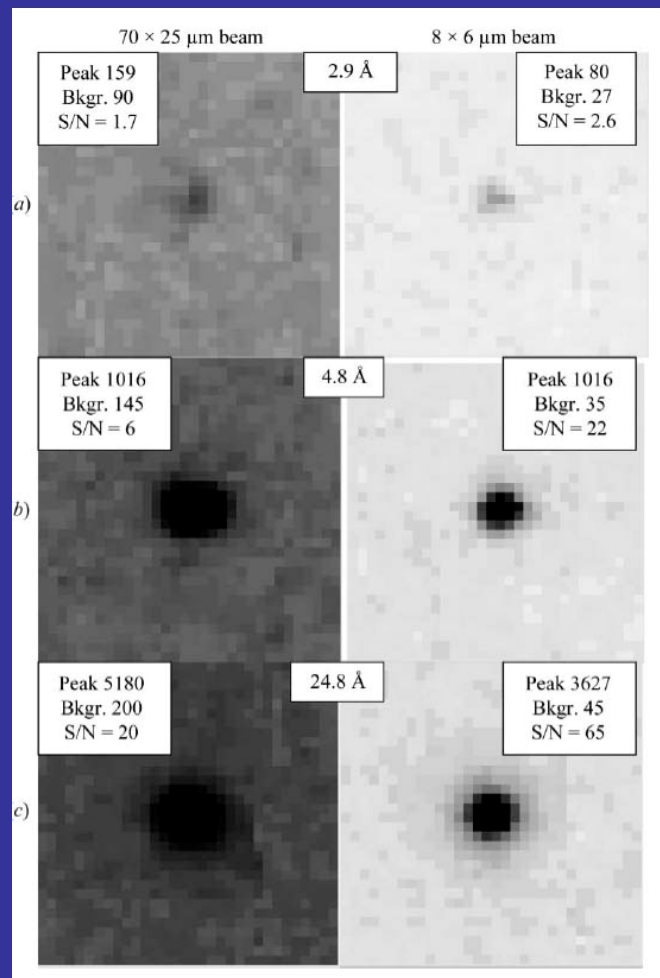
Microbeams improve signal to noise by excluding solvent diffraction

Beam size; 200 μm vs 10 μm





Small vs large beam

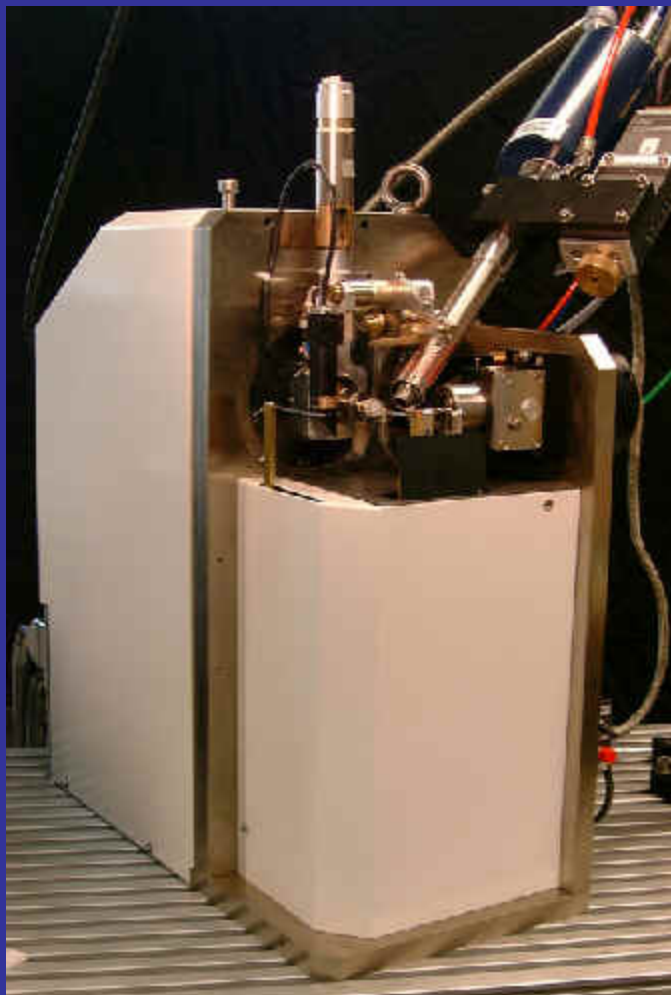


Sanishvili et al, Acta Crys. D 2008

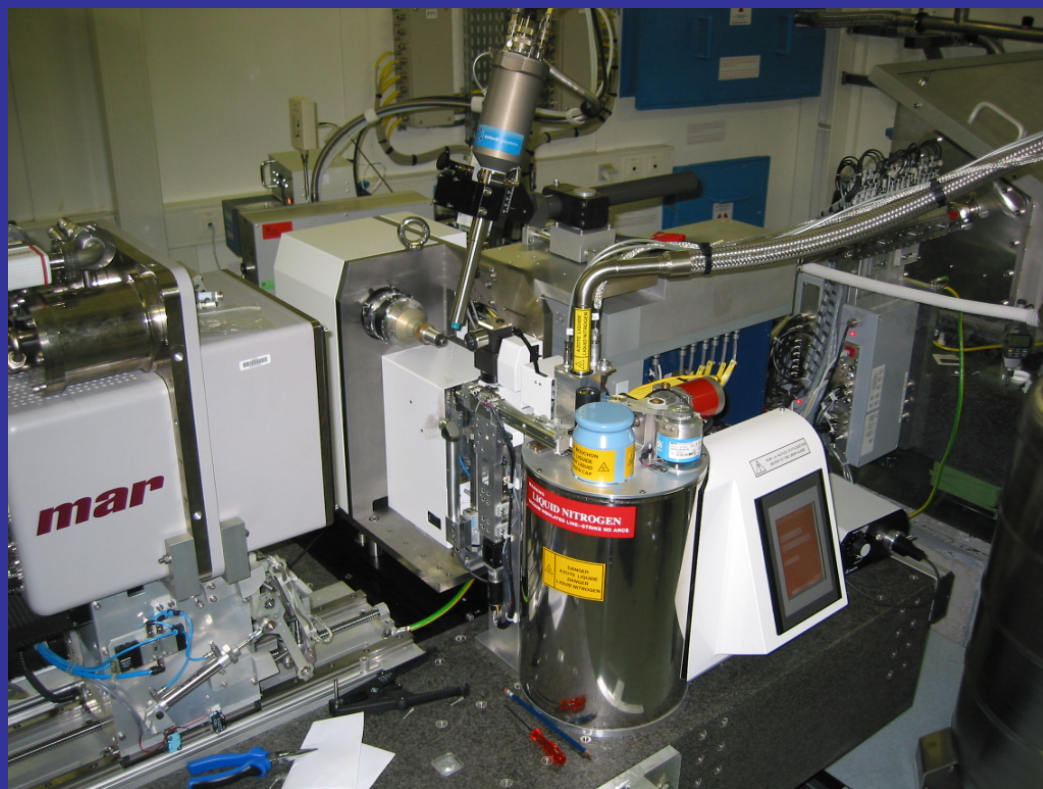


Microfocus beam line set ups

ID13/EMBL

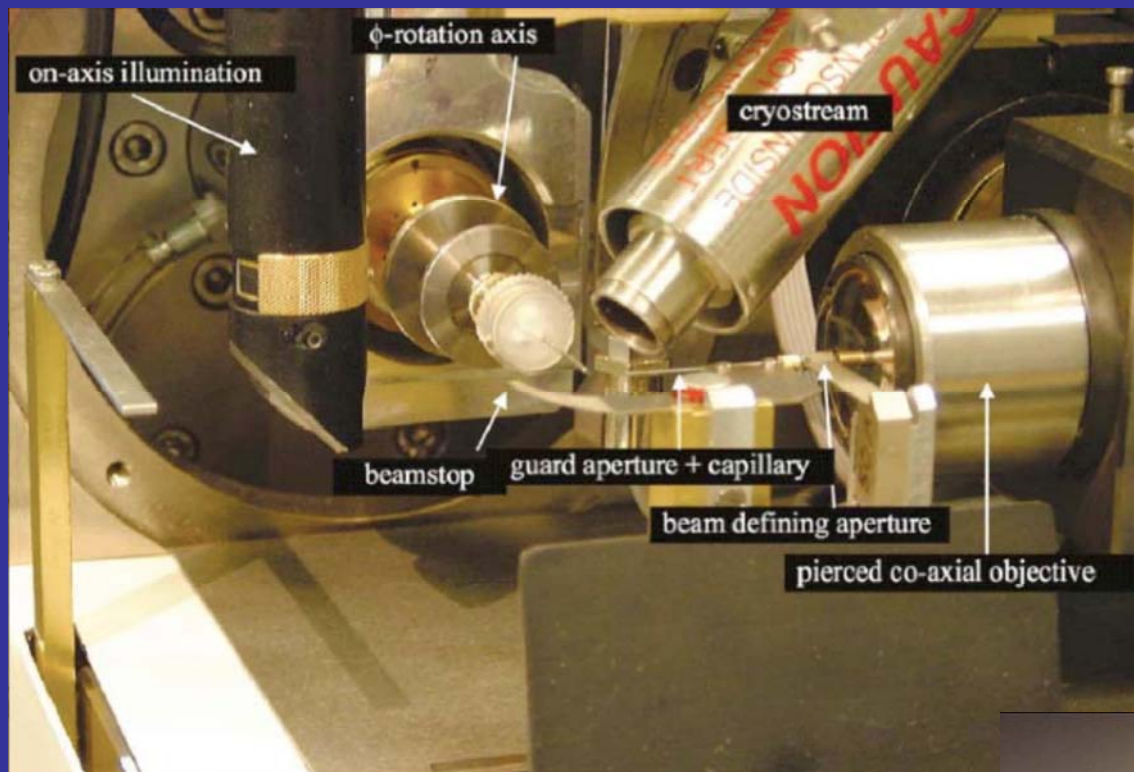


ID23-2

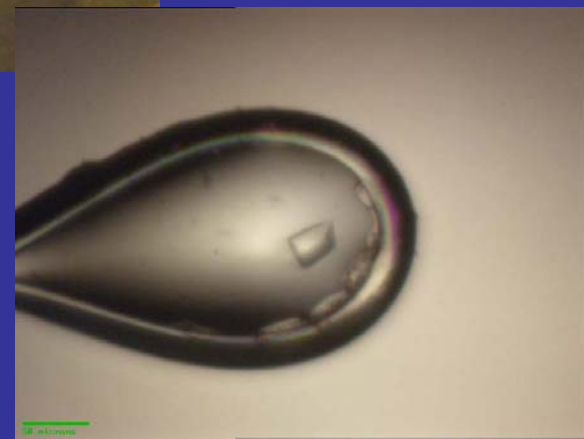




Sample environment

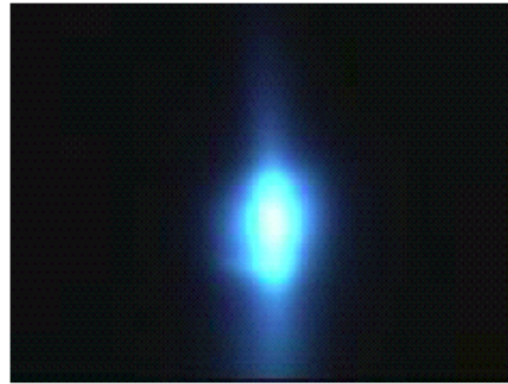


ID13; showing first micro-diffractometer





Beam shaping with guard tube and aperture



**Focussed beam
No shaping**



**100 µm cleaning
aperture**



**30 µm definition
aperture +
100 µm cleaning
aperture**



Detectors

- Mar CCD

ID13, ID23, SLS



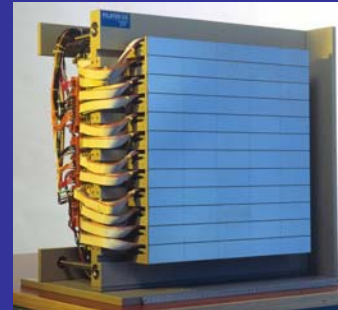
- FreLon

ID13



- Pilatus - Pixel detector

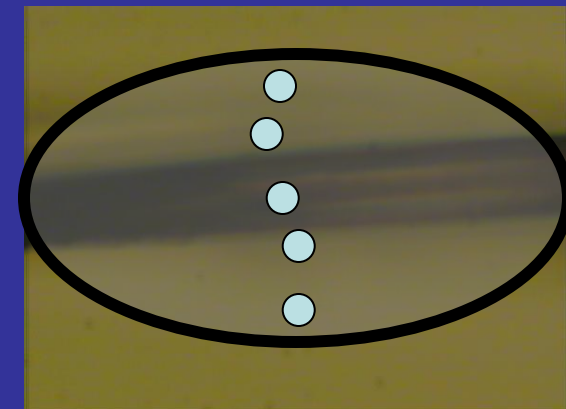
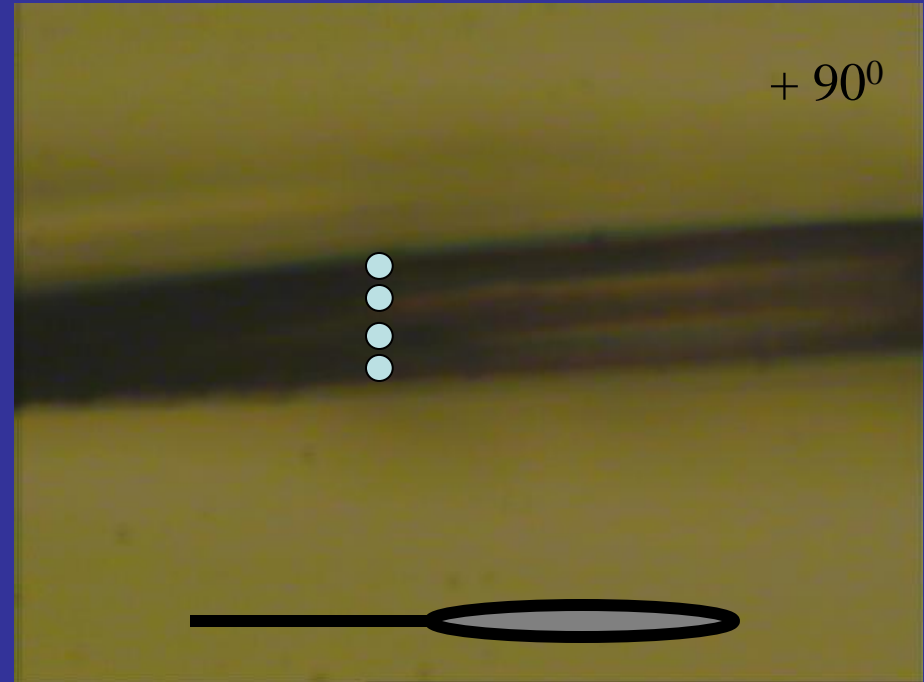
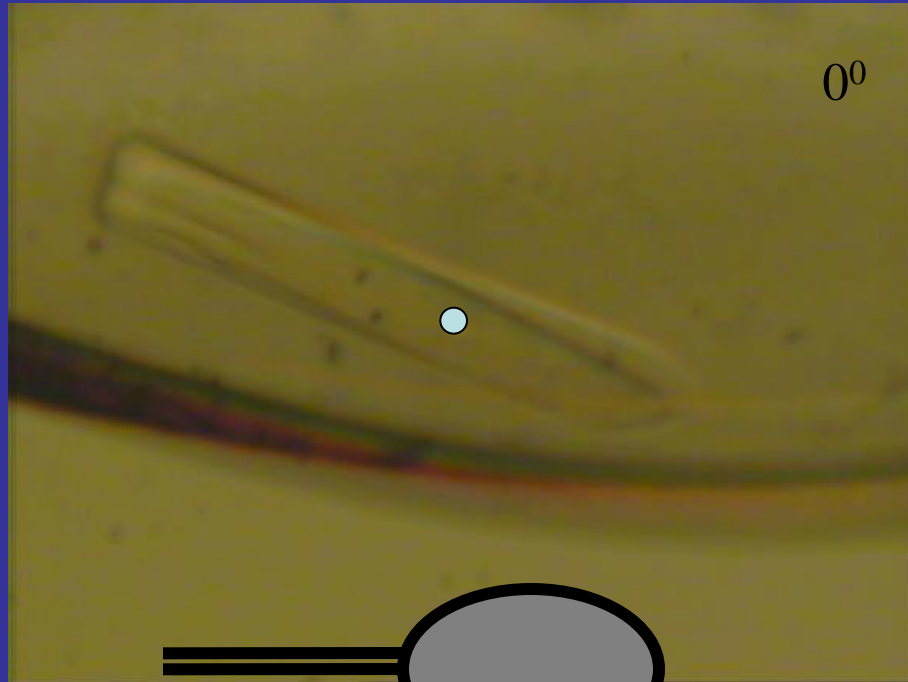
SLS



Important to have fast readout, especially for scanning procedures



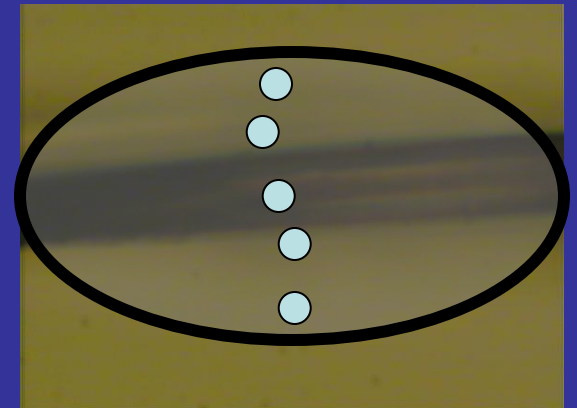
Practicalities- centring crystals is essential with microbeams



Lens effect creates a problem for accurate centring

Sample optimisation

- Cryo buffers tend to create viscous solutions
- Must avoid big fat loops of solution
 - especially in very small $< 0.2\text{mm}$ loops
- Convex solution can causes stress on the needle
- Better to have larger loops
 - with thin film very flat parallel to loop
- Use flatter loops such as LithoLoops or MiTeGen loops



Benefits

- Heterogeneous crystals - choose only best parts
- Many wedge opportunities
- Variable order in different positions leads to variable mosaicity
- Radiation damage; after a limited number of frames can change position
- Multiple crystals can be checked in same loop

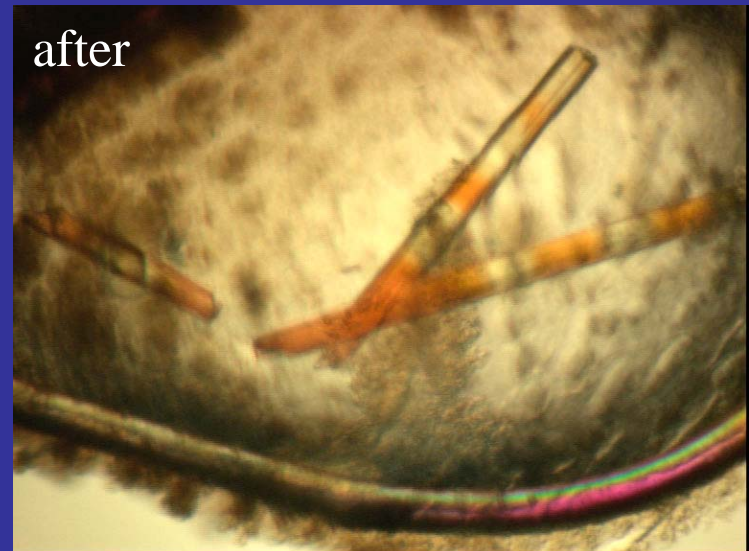
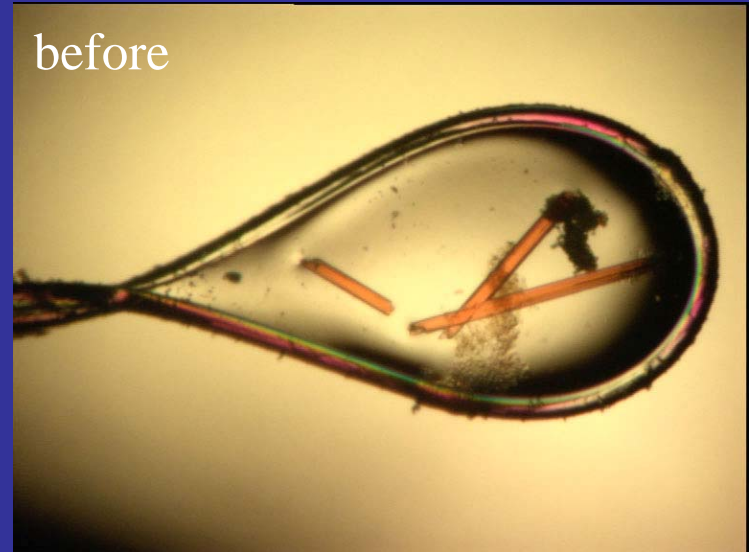
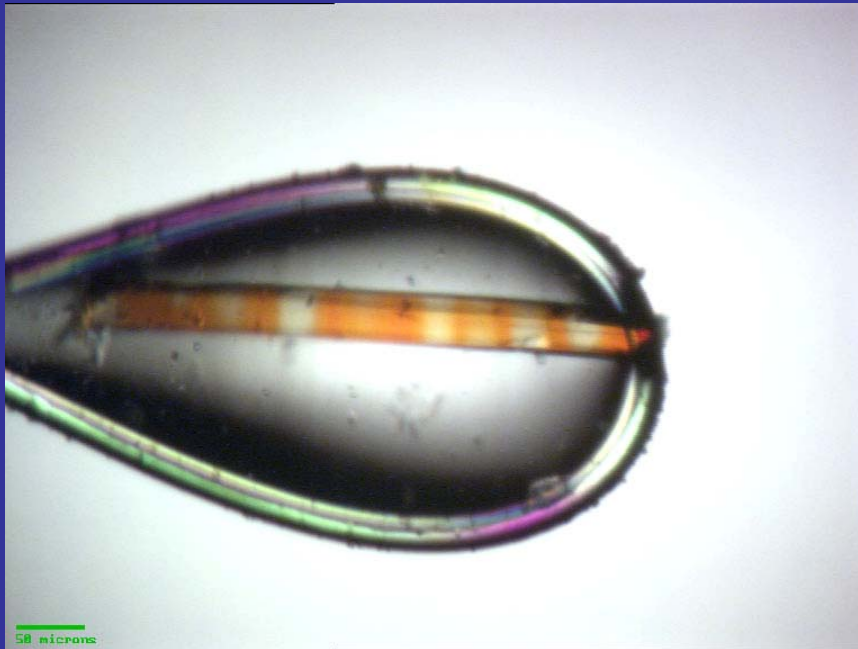
Problems

- Splitting
- Many independent wedges to merge
- Radiation damage
- Data anisotropy - weak diffraction due to misalignment
- Non -isomorphous
- Scaling problems



Localised beam damage

Bovine rhodopsin crystals





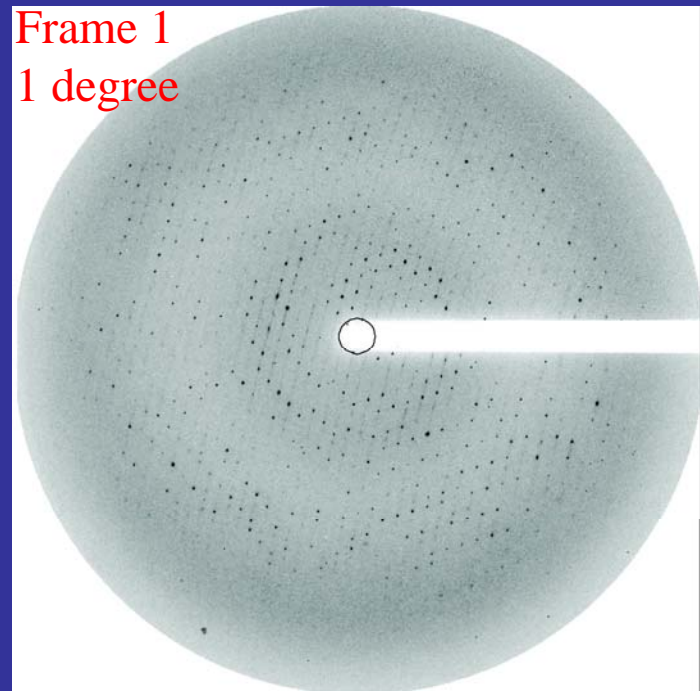
Data collection using ID13 Microfocus beamline

5 micrometer beam on a
5 micrometer recombinant rhodopsin crystal

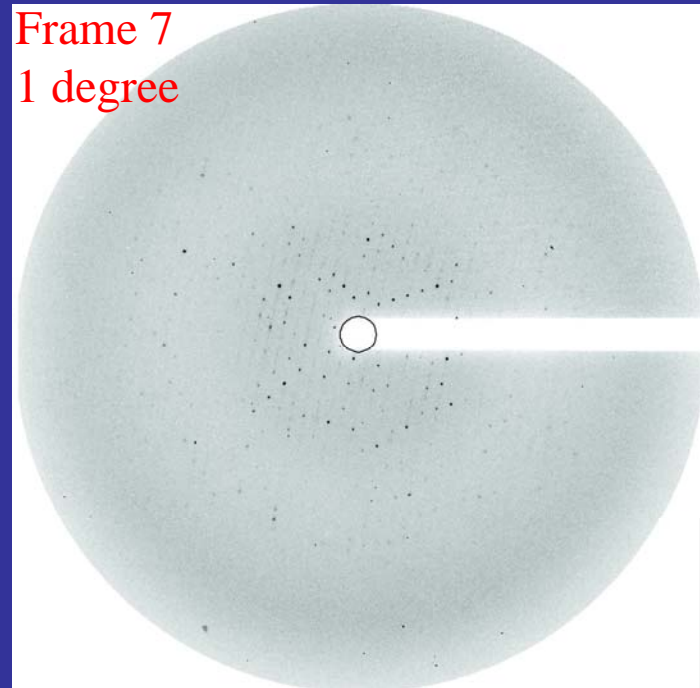


12 data wedges were collected from
Different positions on a single needle

Frame 1
1 degree

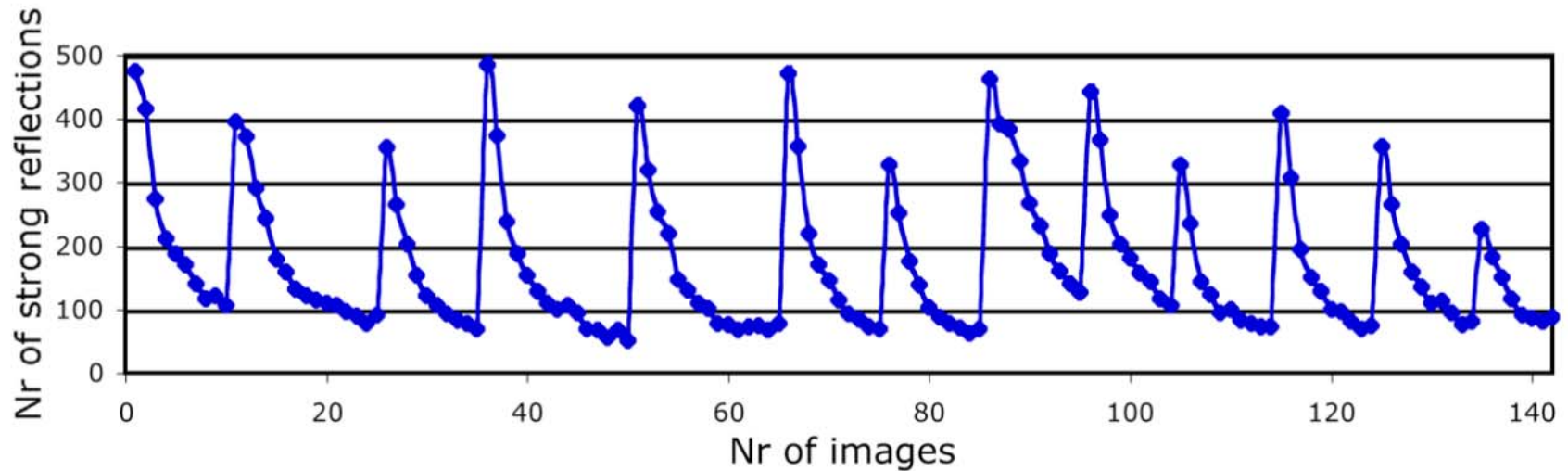


Frame 7
1 degree





Merging of data wedges from different positions

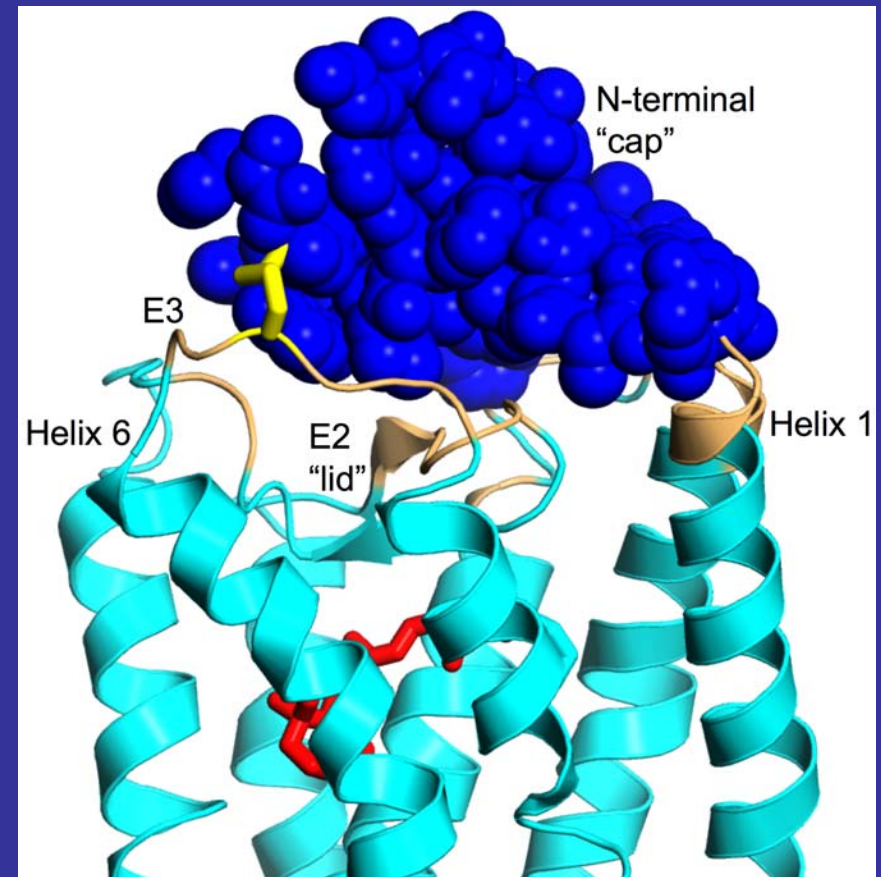
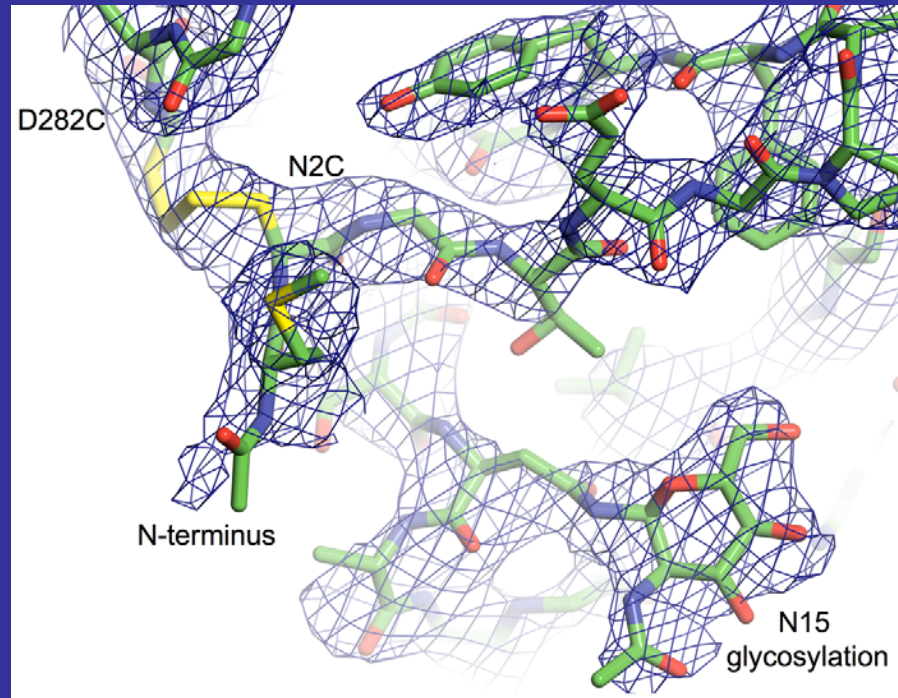


- Merge **as much data as possible** without including **too much radiation damage**
- Find an acceptable compromise

Standfuss et al, JMB 2007



Opsin stabilising disulphide bond fixes the N-terminal cap over the lid



Radiation sensitive disulphide bridge
is visible

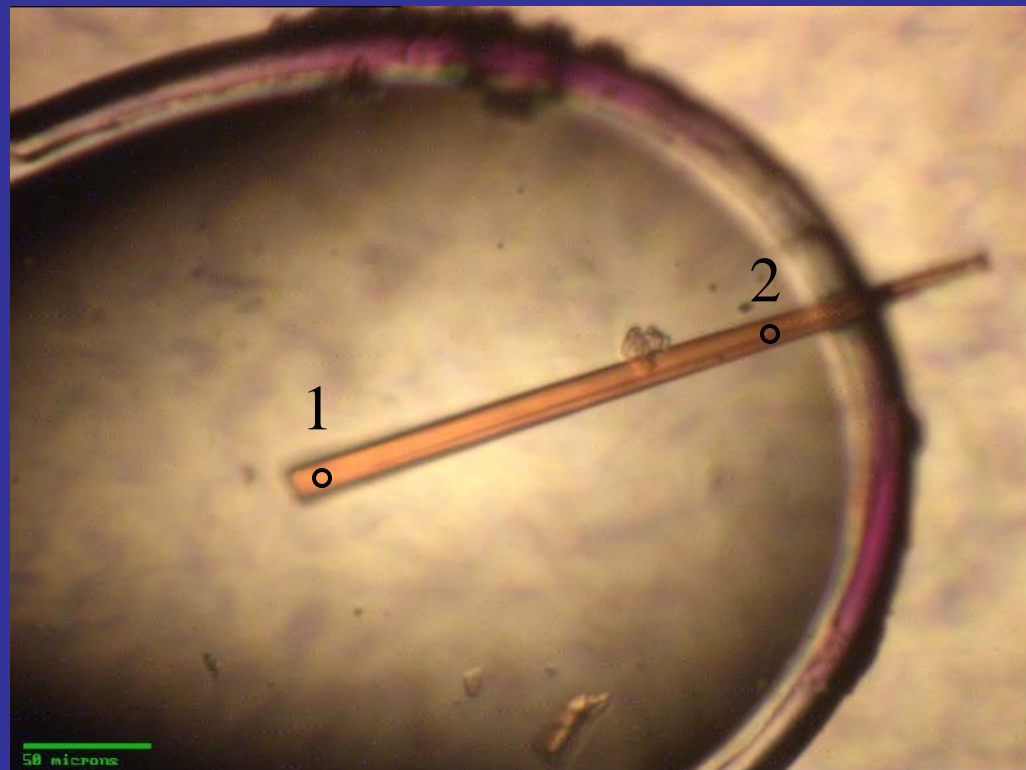
First structure of a recombinant GPCR
Standfuss et al, JMB 2007



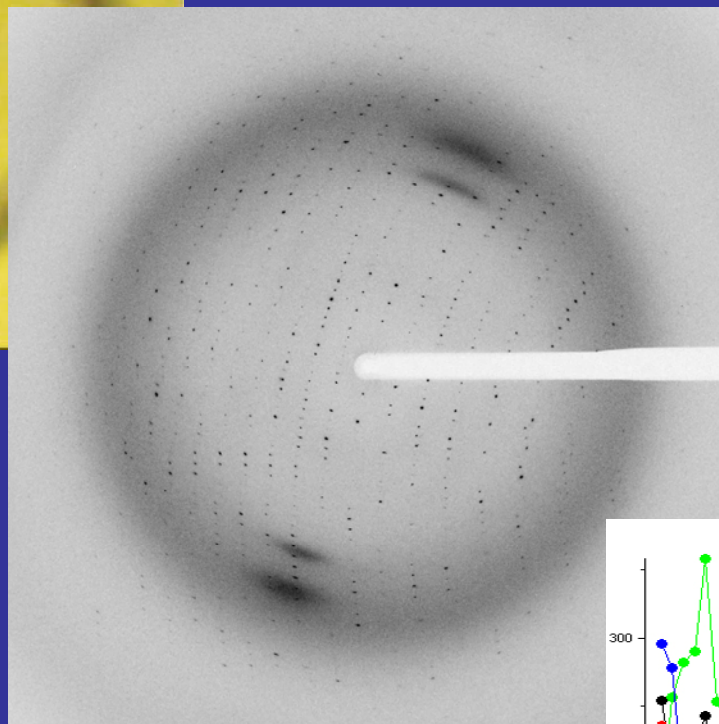
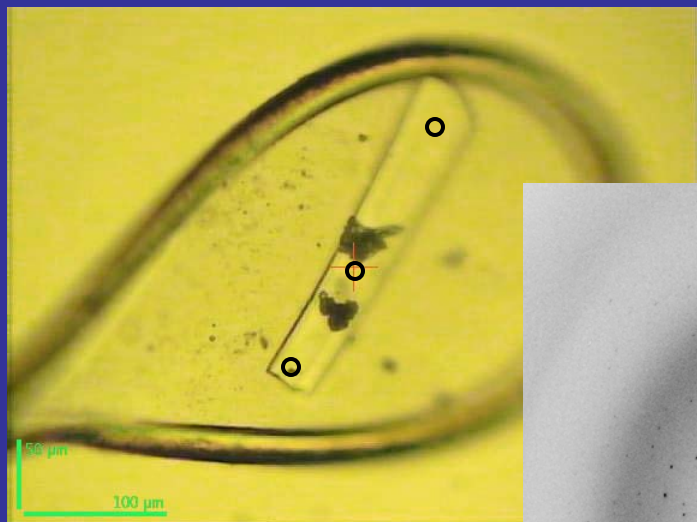
Macros to automate collection

ID23-2- David Flot

Macro for collecting data along a needle - 'helical data collection'.



Beta1 adrenergic receptor; data collection at ID23-2

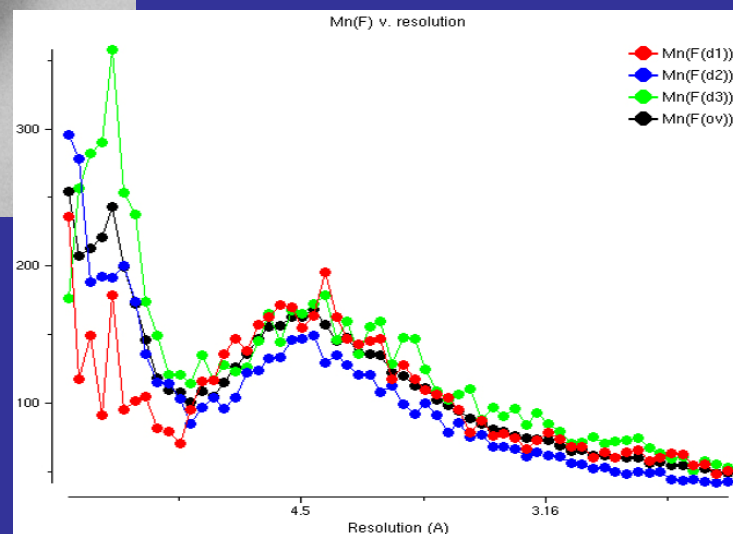


Data isotropic !

Space group P1

Cell parameters: $a=55.5\text{\AA}$, $b=86.8\text{\AA}$, $c=95.50\text{\AA}$

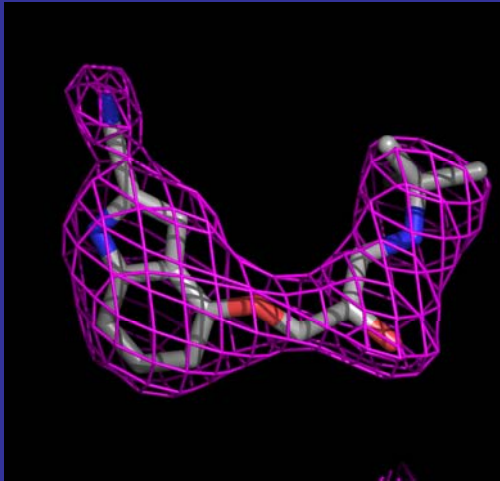
$\alpha=67.60$, $\beta=73.30$, $\gamma=85.80$





Structure of Beta1 adrenergic receptor

Ligand - beta blocker
Cyanopindolol



Warne et al,
Nature 2008

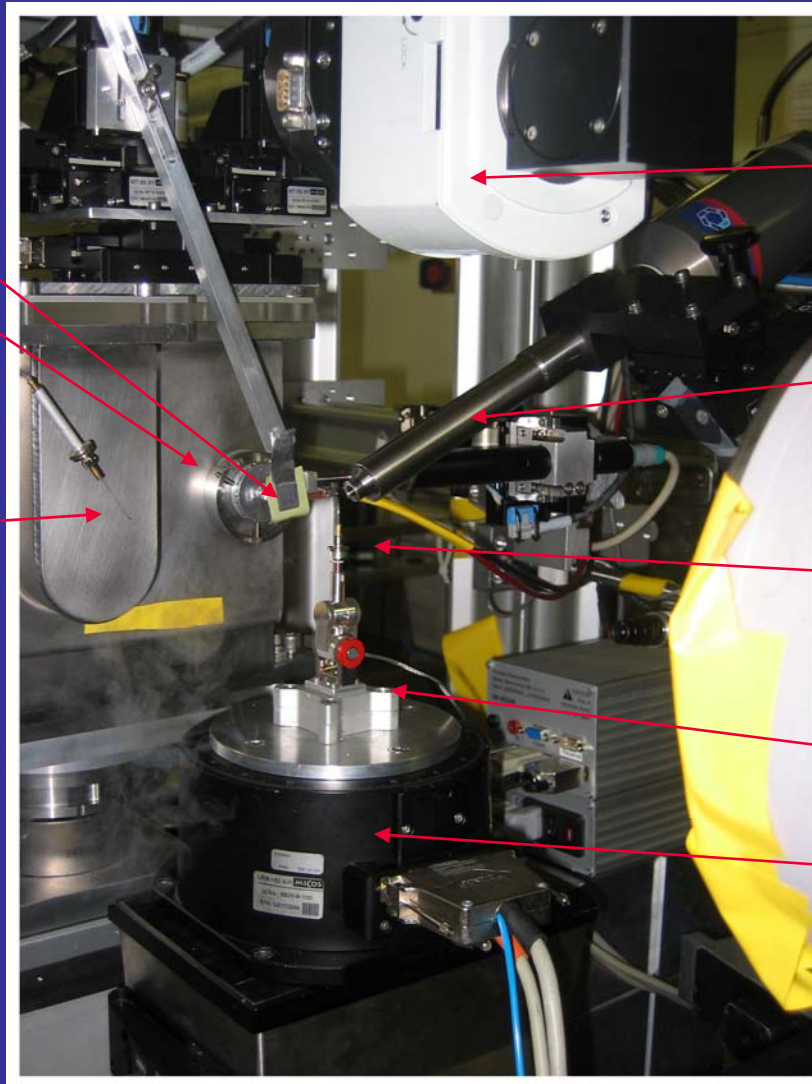
The future...

- Nano beams
- Scanning procedures
- Random data collection



Scanning micro-diffractometer

1 μm beam set up on ID13



Guard aperture support

Mirror chamber

Beamstop

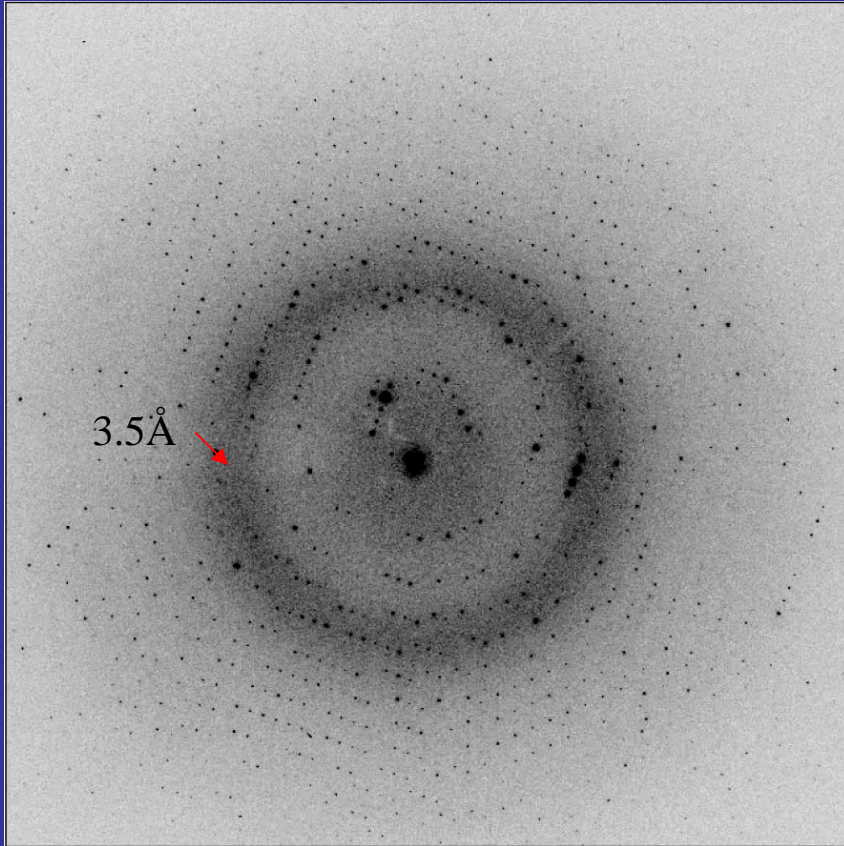
Microscope

Cryostream

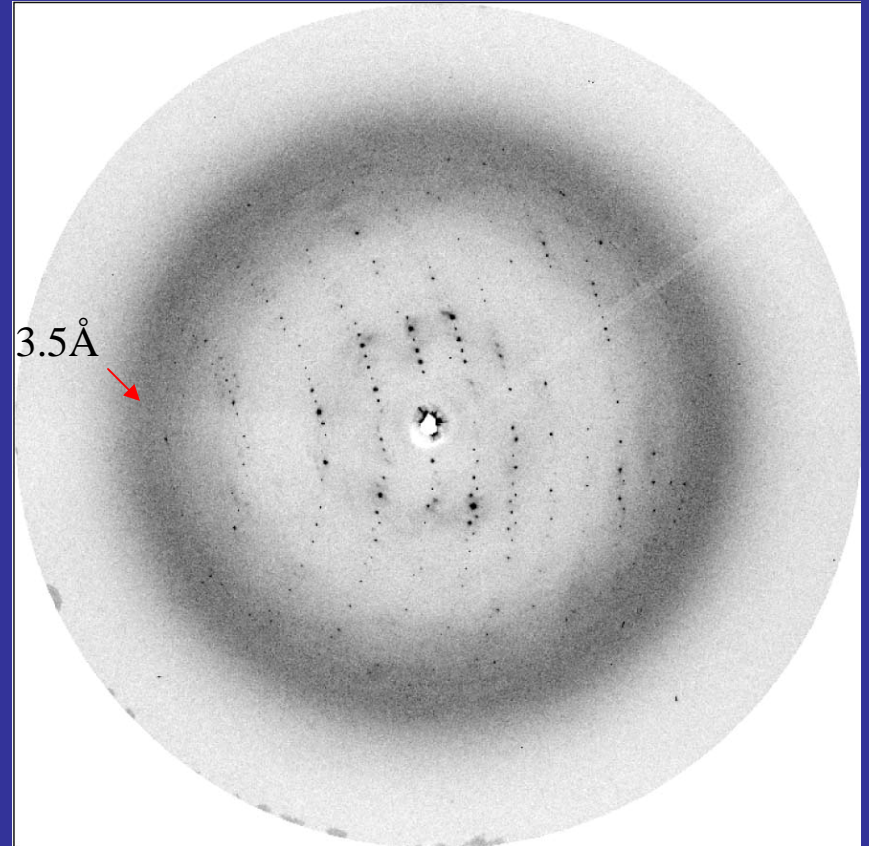
Sample

Piezo micromanipulator
with air bearing rotation axis

1 μ m patterns at ID13



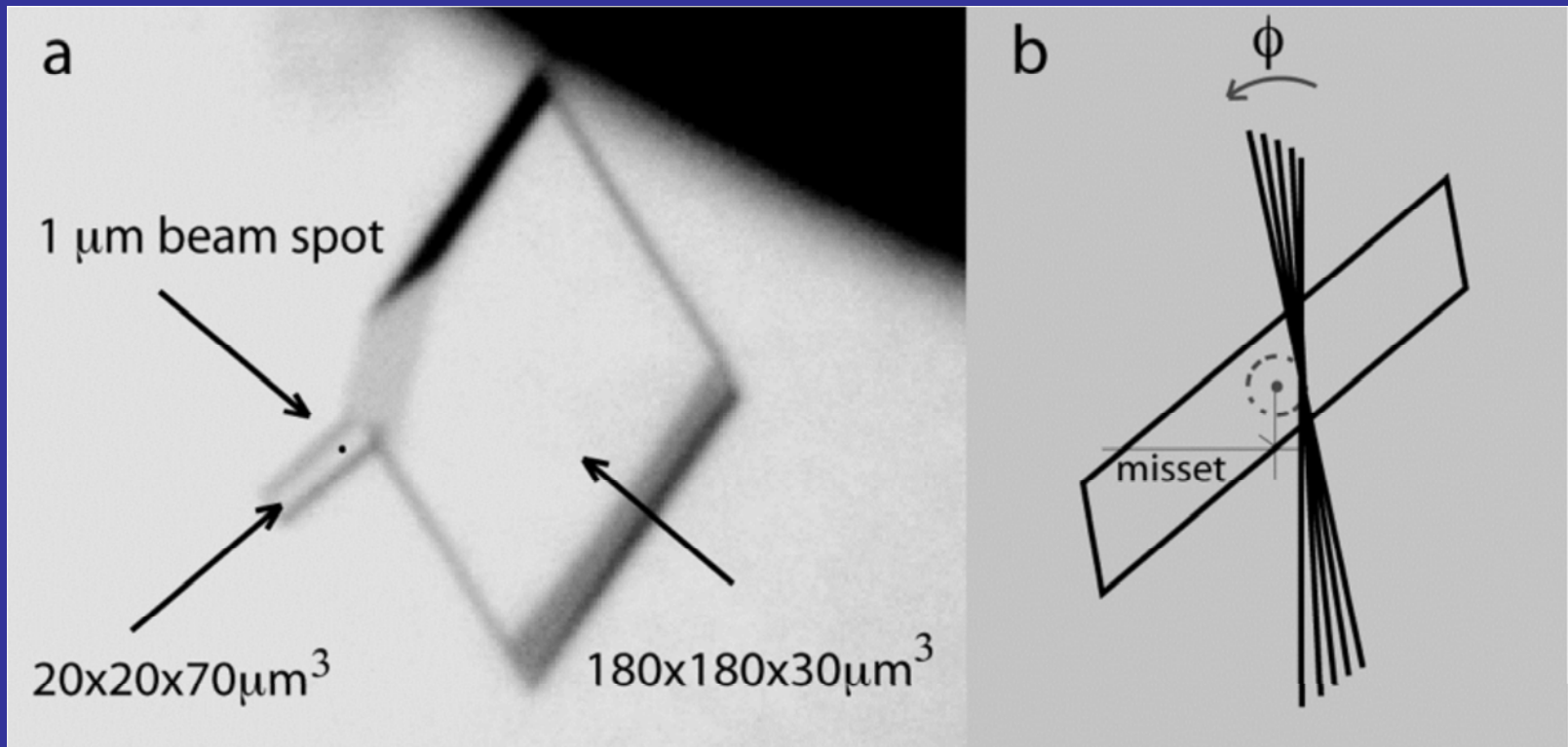
Lysozyme



Beta1 adrenergic receptor



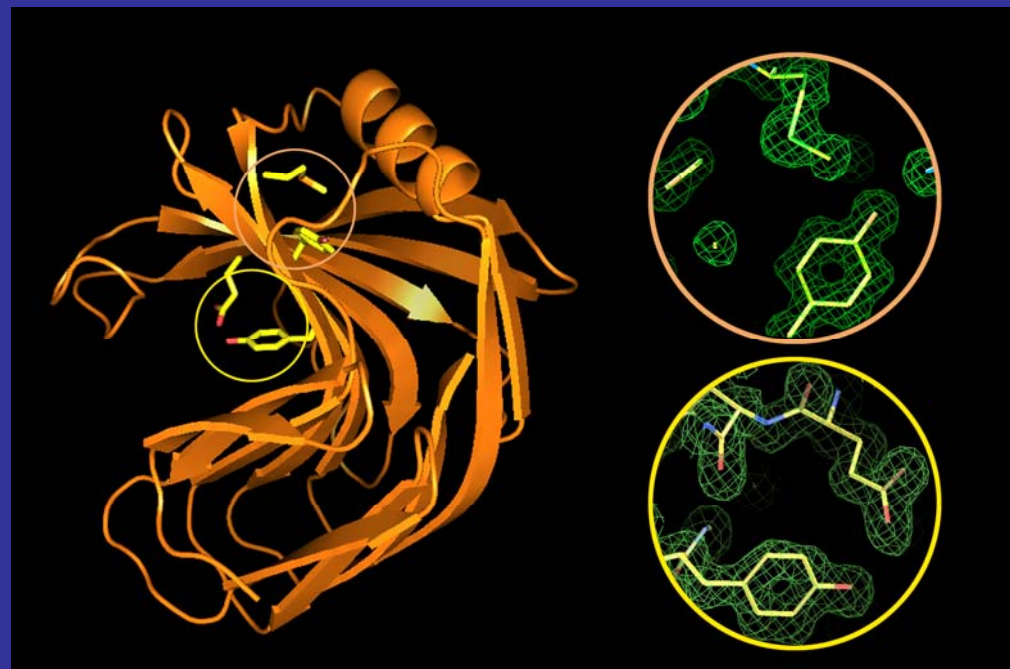
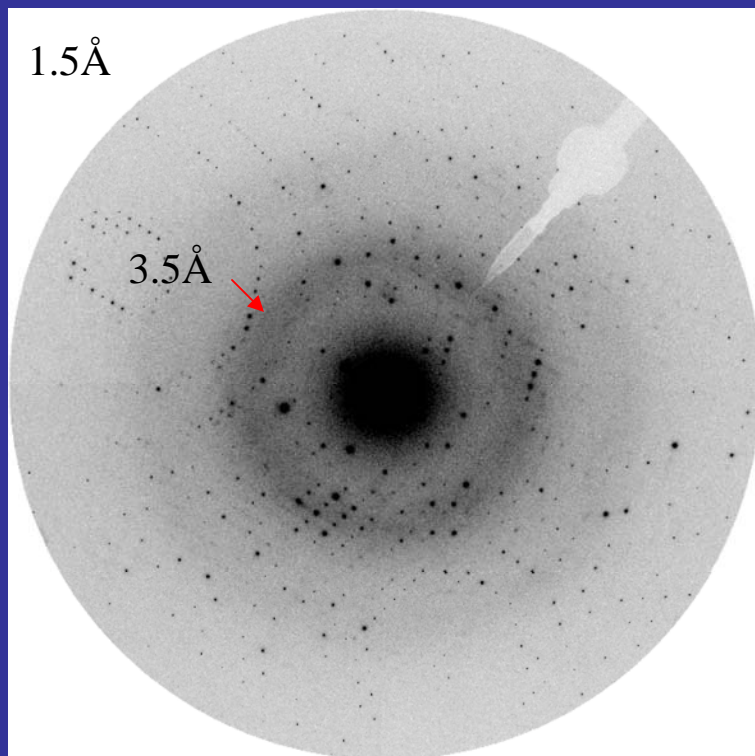
First structure with a $1\mu\text{m}$ X-ray beam at ID13



Xylanase

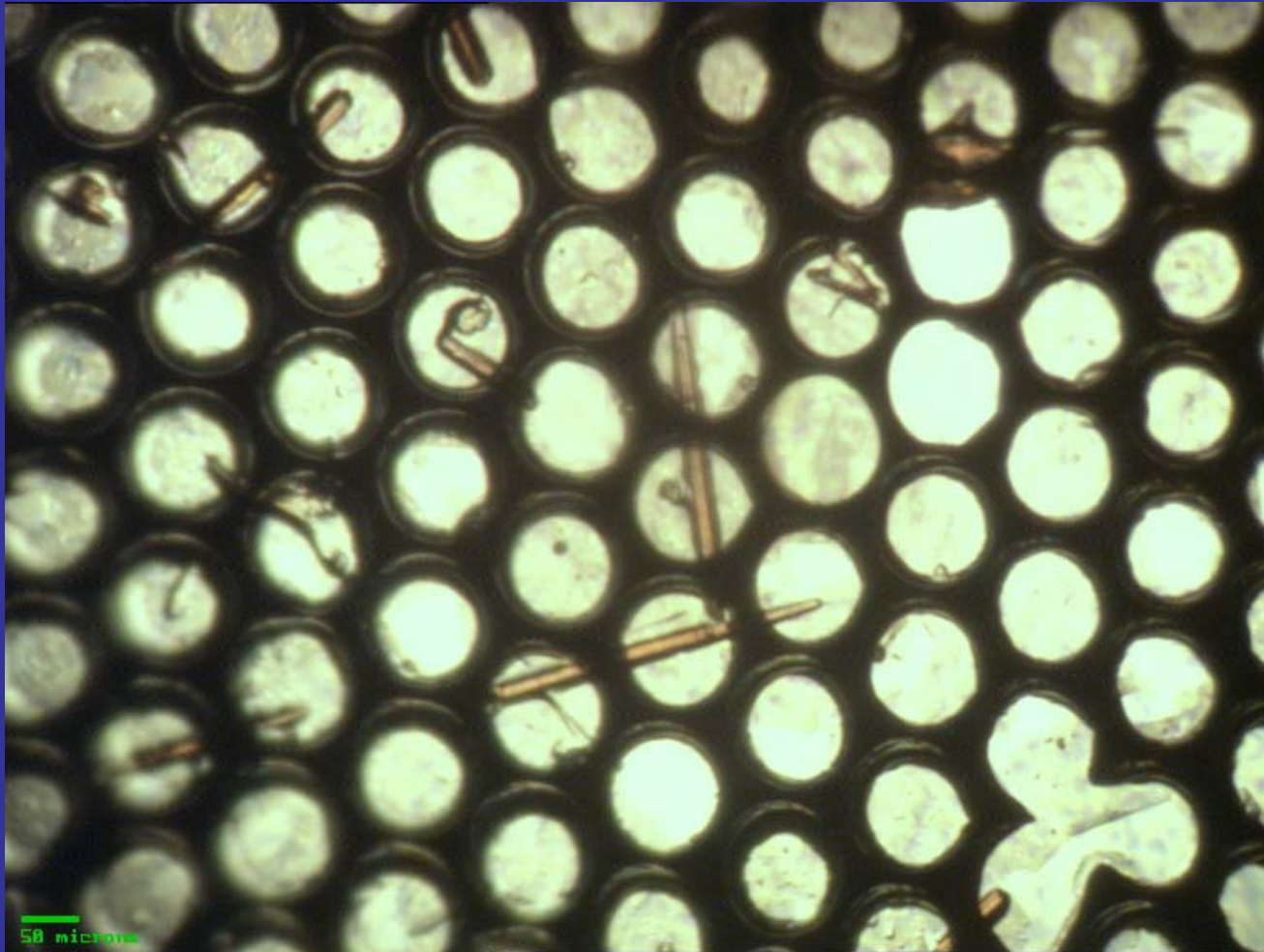


Structure of Xylanase with a 1 μ m beam





Carbon grid mount with rhodopsin crystals



Carbon film support for x-ray diffraction
of protein crystals

Acknowledgements

LMB:

Gebhard Schertler

Jade Li - Bovine rhodopsin

Joerg Standfuss, Dan Oprian - Rhodopsin mutant

Brian Kobilka - Human Beta 2 adrenergic receptor

Tony Warne, Maria Serrano, Chris Tate, Andrew Leslie

- Turkey Beta 1 adrenergic receptor

Rouslan Moukhametzianov, Maikel Fransen - Xylanase

ESRF:

ID13 Manfred Burghammer

ID23-2 David Flot

References

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